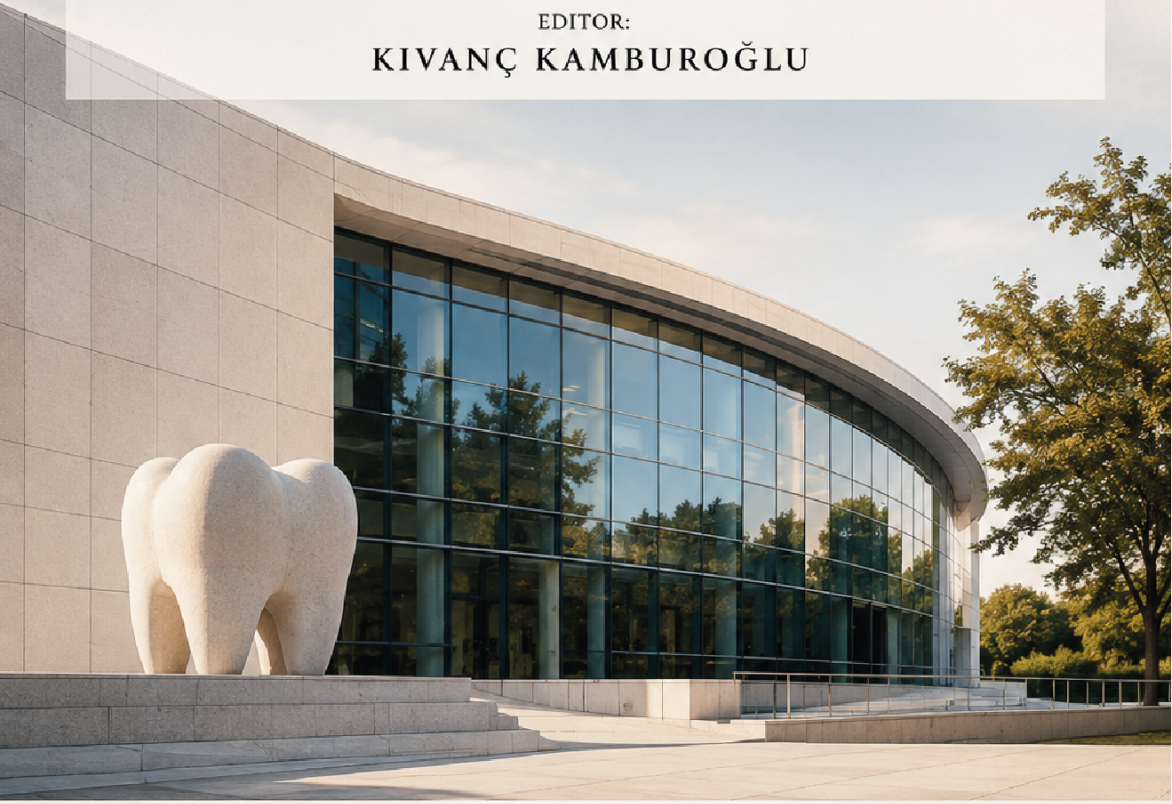


CURRENT ADVANCES

— IN —

ORAL AND MAXILLOFACIAL SCIENCES

EDITOR:
KIVANÇ KAMBUROĞLU



BİDGE Yayınları

Current Advances in Oral and Maxillofacial Sciences

Editor: KIVANÇ KAMBUROGLU

ISBN: 978-625-8989-98-4

1st Edition

Page Layout By: G zde Y CEL

Publication Date: 2026-06-25

BİDGE Yayınları

All rights reserved. No part of this work may be reproduced in any form or by any means, except for brief quotations for promotional purposes with proper source attribution, without the written permission of the publisher and the editor.

Certificate No: 71374

All rights reserved   BİDGE Yayınları

www.bidgeyayinlari.com.tr - bidgeyayinlari@gmail.com

Krc Biliřim Ticaret ve Organizasyon Ltd. řti.

G zeltepe Mahallesi Abidin Daver Sokak Sefer Apartmanı No: 7/9  ankaya /
Ankara



CONTENTS

ORAL CANDIDIASIS IN IMMUNOCOMPROMISED PATIENTS: EMERGING CHALLENGES AND NOVEL THERAPIES	1
---	---

ÖZGE ÖZDAL ZİNCİR

3D PRINTING TECHNOLOGIES IN ORAL AND MAXILLOFACIAL SURGERY	30
---	----

ÖZGE ÖZDAL ZİNCİR

THE ROLE OF VITAMIN D IN BONE METABOLISM: FROM MOLECULAR FOUNDATIONS TO THERAPEUTIC APPLICATIONS IN ORAL SURGERY	58
--	----

İSMAİL FARUK OKUMUŞ, GÖKHAN GÜRSES

GRAFT MATERIALS AND MECHANISMS OF BONE FORMATION	73
---	----

GÖKHAN GÜRSES, İSMAİL FARUK OKUMUŞ

CHAPTER 1

Oral Candidiasis in Immunocompromised Patients: Emerging Challenges and Novel Therapies

Özge ÖZDAL ZİNCİR¹

Introduction

Oral candidiasis is one of the most common opportunistic fungal infections affecting the oral cavity and represents a major clinical challenge particularly in immunocompromised individuals. Although *Candida* species are normal commensal microorganisms within the oral microbiota, alterations in host immunity, oral environment, or microbial balance may facilitate pathogenic transformation and fungal overgrowth (Williams & Lewis, 2000).

Among *Candida* species, *Candida albicans* remains the most frequently isolated pathogen in oral candidiasis. However, non-*albicans* *Candida* species such as *Candida glabrata*, *Candida tropicalis*, *Candida krusei*, and *Candida parapsilosis* have increasingly emerged in immunocompromised populations and are often associated with antifungal resistance and recurrent infections (Silva, et al., 2012).

¹ Assoc. Prof. Dr. Istanbul University- Cerrahpasa, Faculty of Dentistry, Department of Oral and Maxillofacial Surgery ORCID: 0009-0000-9423-5764

The prevalence of oral candidiasis has increased substantially over recent decades because of the growing number of immunocompromised patients. Individuals with HIV/AIDS, hematological malignancies, solid organ transplantation, chemotherapy-induced immunosuppression, uncontrolled diabetes mellitus, corticosteroid therapy, autoimmune diseases, malnutrition, and advanced age are particularly susceptible to fungal colonization and infection (Samaranayake, 1992).

Oral candidiasis may significantly impair quality of life by causing pain, dysphagia, dysgeusia, burning sensation, nutritional difficulties, and reduced oral function. In severely immunocompromised patients, oral *Candida* infections may additionally serve as reservoirs for systemic dissemination, potentially leading to life-threatening invasive candidiasis (Calderone & Clancy, 2012).

The pathogenesis of oral candidiasis is multifactorial and involves complex interactions between host immune defenses, fungal virulence factors, salivary components, microbiome alterations, and local environmental conditions (Mayer, Wilson & Hube, 2013). *Candida* species possess numerous virulence mechanisms including biofilm formation, phenotypic switching, adhesion molecules, hydrolytic enzyme secretion, and immune evasion capabilities.

One of the major contemporary concerns is the increasing incidence of antifungal resistance. Prolonged or repeated antifungal exposure, particularly azole therapy, has contributed to the emergence of resistant *Candida* strains (Perfect, 2017). Biofilm-associated infections further complicate treatment because biofilms demonstrate significantly increased resistance compared with planktonic fungal cells.

Conventional antifungal therapies such as nystatin, clotrimazole, fluconazole, and amphotericin B remain widely used; however, therapeutic limitations including resistance, recurrence, adverse effects, and limited biofilm penetration continue to challenge clinicians (Pappas, et al., 2016). Consequently, considerable research has focused on development of novel antifungal approaches including photodynamic therapy, photobiomodulation, nanotechnology-based systems, probiotics, antimicrobial peptides, natural compounds, and combination therapies.

Recent advances in molecular biology, immunology, microbiome analysis, and fungal genomics have significantly improved understanding of oral candidiasis pathogenesis and therapeutic targets (Brown, et al., 2012). Novel translational strategies integrating antifungal agents with immunomodulatory and biofilm-targeting therapies are increasingly being investigated.

This chapter reviews the epidemiology, pathogenesis, clinical manifestations, diagnostic approaches, antifungal resistance mechanisms, and emerging therapeutic strategies for oral candidiasis in immunocompromised patients.

Candida Species and Oral Microbiota

Candida species are opportunistic fungal organisms that commonly colonize the oral cavity of healthy individuals without causing disease. Approximately 30–60% of healthy adults may harbor *Candida* organisms within the oral microbiome (Akpan & Morgan, 2002). Colonization rates increase significantly in hospitalized, elderly, denture-wearing, and immunocompromised populations.

Candida albicans remains the predominant species associated with oral candidiasis because of its remarkable adaptability and virulence capacity (Calderone, 2002). Nevertheless, the

epidemiological importance of non-albicans *Candida* species has increased substantially over recent years.

Candida glabrata has become particularly important in immunocompromised individuals because of its intrinsic reduced susceptibility to azole antifungal agents (Fidel, Vazquez & Sobel, 1999). Unlike *C. albicans*, *C. glabrata* does not form true hyphae; however, it demonstrates strong adhesive properties and biofilm formation capabilities.

Candida tropicalis and *Candida parapsilosis* are additionally important opportunistic pathogens, especially among oncology and intensive care patients (Pfaller & Diekema, 2007). *Candida krusei* is clinically significant because of its intrinsic resistance to fluconazole.

The oral microbiome plays an important role in regulating fungal colonization. Commensal bacteria may inhibit *Candida* overgrowth through competitive interactions and antimicrobial metabolite production (Jenkinson & Lamont, 2005). Disruption of microbial balance by antibiotics, xerostomia, poor oral hygiene, or immunosuppression may facilitate fungal proliferation.

Saliva additionally serves as an important defense mechanism against fungal colonization. Salivary proteins including histatins, defensins, lactoferrin, lysozyme, calprotectin, and secretory immunoglobulin A possess antifungal activity (Edgerton & Koshlukova, 2000). Reduced salivary flow or altered salivary composition significantly increases candidiasis susceptibility.

The transition of *Candida* species from harmless commensals to pathogenic organisms depends largely on host immune dysfunction and environmental changes. Understanding these host-microbe interactions remains essential for development of effective preventive and therapeutic strategies.

Immunocompromised Conditions Associated with Oral Candidiasis

Immunocompromised patients represent the population at greatest risk for oral candidiasis. Impaired immune surveillance allows fungal overgrowth and increases susceptibility to recurrent and severe infections (Williams & Lewis, 2011).

HIV/AIDS historically represented one of the most important risk factors for oral candidiasis. Before widespread antiretroviral therapy, oral candidiasis was among the earliest and most common clinical manifestations of HIV infection (Greenspan, 1997). Reduced CD4+ T-cell counts significantly impair mucosal immunity and facilitate *Candida* invasion.

Cancer patients receiving chemotherapy or radiotherapy are also highly susceptible to oral fungal infections. Cytotoxic therapies may cause mucosal injury, neutropenia, xerostomia, and microbiome disruption, all of which favor *Candida* colonization (Sonis, 2004). Oral mucositis additionally creates an environment conducive to fungal invasion.

Hematopoietic stem cell transplantation and solid organ transplantation are associated with prolonged immunosuppressive therapy and increased opportunistic infection risk (Imanguli, et al., 2008). Corticosteroids, calcineurin inhibitors, and other immunosuppressive agents impair both innate and adaptive immune responses.

Diabetes mellitus is another major predisposing factor. Hyperglycemia, impaired neutrophil function, reduced salivary flow, and altered oral microbiota contribute significantly to increased *Candida* colonization in diabetic patients (Guggenheimer, et al., 2000).

Elderly individuals frequently demonstrate multiple risk factors including xerostomia, denture use, malnutrition, polypharmacy, and age-related immune decline (Coco, et al., 2008). Consequently, oral candidiasis prevalence increases substantially in geriatric populations.

Autoimmune diseases such as Sjögren syndrome may predispose patients to candidiasis because of severe salivary dysfunction and mucosal dryness (Fox, 2005). Similarly, prolonged corticosteroid inhaler use in asthma patients may promote localized fungal overgrowth.

Malnutrition, smoking, alcohol abuse, intensive care hospitalization, and prolonged antibiotic therapy additionally contribute to increased fungal susceptibility (Soysa & Ellepola, 2005). The combination of multiple predisposing factors often results in chronic or recurrent oral candidiasis that is difficult to manage.

Pathogenesis and Virulence Factors

The pathogenesis of oral candidiasis involves complex interactions between fungal virulence mechanisms and host immune defenses. *Candida* species possess numerous adaptive strategies that facilitate adhesion, tissue invasion, immune evasion, and persistence (Hube, 2004).

Adhesion to epithelial surfaces represents the initial step in colonization. *Candida* species express adhesins and cell surface proteins that enable attachment to oral mucosa, dentures, and biomaterial surfaces (Sundstrom, 2002). This adhesion process is essential for biofilm formation and infection establishment.

Morphological switching is another major virulence factor, particularly in *Candida albicans*. Transition from yeast form to filamentous hyphal form enhances tissue invasion and pathogenicity

(Sudbery, Gow & Berman, 2004). Hyphal forms may penetrate epithelial tissues and evade phagocytic defenses more effectively than yeast cells.

Biofilm formation represents one of the most clinically important pathogenic mechanisms. *Candida* biofilms are highly organized microbial communities embedded within extracellular polymeric matrices (Ramage, Martínez & López-Ribot, 2006). Biofilms frequently develop on dentures, implants, mucosal surfaces, and medical devices.

Biofilm-associated *Candida* cells demonstrate significantly increased resistance to antifungal agents and host immune defenses (Nett & Andes, 2020). Reduced drug penetration, altered metabolic activity, stress-response activation, and extracellular matrix protection all contribute to enhanced resistance.

Candida species additionally secrete hydrolytic enzymes including secreted aspartyl proteinases, phospholipases, and lipases (Naglik, Challacombe & Hube, 2003). These enzymes facilitate tissue invasion, epithelial damage, nutrient acquisition, and immune evasion.

Host immune responses play a central role in controlling fungal colonization. Neutrophils, macrophages, dendritic cells, and T-helper 17 lymphocytes are particularly important in antifungal immunity (Netea, et al., 2015). Impairment of these immune pathways significantly increases susceptibility to candidiasis.

Inflammatory cytokines including IL-1 β , IL-6, TNF- α , and IL-17 contribute to mucosal defense against *Candida* infection (Gladiator, et al., 2013). However, excessive inflammatory responses may additionally contribute to tissue damage and symptomatic disease.

Recent research has also emphasized the importance of microbiome interactions in fungal pathogenicity. Bacterial-fungal synergy may influence biofilm formation, antifungal resistance, and disease severity (Xu, Jenkinson & Dongari-Bagtzoglou, 2014). Understanding these polymicrobial interactions may provide novel therapeutic opportunities in the future.

Clinical Manifestations of Oral Candidiasis

Oral candidiasis may present with diverse clinical manifestations depending on host immunity, *Candida* species involved, and local environmental factors. In immunocompromised patients, lesions are often more extensive, recurrent, and resistant to conventional therapy (Farah, Lynch & McCullough, 2010).

Pseudomembranous candidiasis, commonly referred to as thrush, is the most recognizable clinical form. It is characterized by white or yellowish removable plaques on the oral mucosa, tongue, palate, and oropharynx (Samaranayake & Holmstrup, 1989). Removal of the pseudomembrane often reveals erythematous or bleeding mucosa underneath. This form is particularly common in HIV-positive patients, chemotherapy recipients, and individuals receiving corticosteroids.

Erythematous candidiasis presents as red, painful, and atrophic mucosal lesions, frequently affecting the tongue and palate (Budtz-Jorgensen, 1990). Patients often report burning sensation, dysgeusia, and oral discomfort. This subtype is commonly associated with broad-spectrum antibiotic use and xerostomia.

Hyperplastic candidiasis is less common but clinically significant because it may mimic leukoplakia and possesses potential associations with epithelial dysplasia (Sitheeque & Samaranayake, 2003). Lesions typically appear as non-removable white plaques, most commonly located at the commissural or buccal mucosa.

Angular cheilitis represents another frequent manifestation characterized by erythematous fissuring at the corners of the mouth (Sharon & Fazel, 2010). Candida infection often occurs in combination with bacterial colonization and is particularly common in elderly denture wearers.

Denture stomatitis is one of the most prevalent Candida-associated conditions among removable prosthesis users. Poor denture hygiene, continuous denture wearing, trauma, and biofilm accumulation contribute significantly to disease development (Webb, et al., 1998). Lesions usually present as diffuse erythema beneath the denture-bearing mucosa.

Median rhomboid glossitis and chronic multifocal candidiasis are additional less common manifestations observed in immunocompromised populations (Reamy, Derby & Bunt, 2010). In severe cases, Candida infection may extend into the oropharynx and esophagus, leading to dysphagia and odynophagia.

Symptoms vary substantially among patients and may include pain, burning sensation, altered taste perception, xerostomia, halitosis, and difficulties in eating or speaking (Gonsalves, Chi & Neville, 2007). Such symptoms may significantly impair nutritional status and quality of life, particularly in oncology and HIV patients.

Because clinical presentations may mimic other oral mucosal diseases, accurate diagnosis requires careful clinical evaluation combined with microbiological or histopathological confirmation in selected cases.

Diagnostic Approaches

Diagnosis of oral candidiasis is primarily based on clinical examination; however, microbiological and molecular diagnostic methods may be necessary in atypical, recurrent, or resistant cases (Coronado-Castellote & Jiménez-Soriano, 2013).

Clinical diagnosis often relies on characteristic lesion appearance and response to antifungal therapy. Nevertheless, differentiation from leukoplakia, lichen planus, geographic tongue, frictional keratosis, and mucosal dysplasia may sometimes be difficult (Neville, et al., 2016).

Cytological examination using exfoliative smears remains a widely used diagnostic method. Potassium hydroxide preparation, Gram staining, and periodic acid-Schiff staining may reveal fungal hyphae or yeast cells (Arendorf & Walker, 1980). Presence of hyphal forms is generally considered indicative of active infection rather than simple colonization.

Culture techniques are useful for species identification and antifungal susceptibility testing. Sabouraud dextrose agar and chromogenic media may facilitate differentiation of *Candida* species (Odds, 1988). Identification of non-albicans *Candida* species is particularly important because of varying antifungal susceptibility patterns.

Molecular diagnostic methods such as polymerase chain reaction and sequencing technologies have significantly improved sensitivity and specificity (Kanbe, et al., 2002). These techniques may allow rapid detection of *Candida* species and resistance-associated genetic mutations.

Salivary diagnostics have additionally emerged as promising noninvasive tools in candidiasis evaluation. Salivary cytokines, oxidative stress markers, fungal metabolites, and microbiome profiles may provide valuable information regarding disease activity and therapeutic response (Ellepola & Samaranayake, 2000).

Biofilm analysis has become increasingly important in denture-related and implant-associated candidiasis. Advanced microscopy and molecular approaches may improve understanding

of polymicrobial interactions and resistance mechanisms (Ramage, et al., 2004).

Histopathological examination may occasionally be necessary in chronic hyperplastic lesions to exclude epithelial dysplasia or malignancy (Eversole, 2001). Tissue invasion by fungal hyphae supports the diagnosis of active infection.

Artificial intelligence-assisted diagnostic systems and machine learning-based image analysis represent emerging fields that may improve future diagnostic precision and screening efficiency (Schwendicke, et al., 2019).

Conventional Antifungal Therapies

Conventional antifungal treatment remains the cornerstone of oral candidiasis management. Therapeutic selection depends on disease severity, patient immunity, fungal species, and recurrence risk (Epstein & Polsky, 1998).

Topical antifungal agents are commonly used in mild or localized infections. Nystatin remains one of the most widely prescribed topical agents because of its broad antifungal spectrum and low systemic toxicity (Lyu, et al., 2016). It is typically administered as oral suspension, cream, or pastille formulations.

Clotrimazole troches are another frequently used topical therapy. These agents provide prolonged mucosal contact and are generally effective in uncomplicated candidiasis (Pien, et al., 1991). However, patient compliance may be limited because of frequent dosing requirements.

Systemic antifungal therapy is often necessary in moderate-to-severe infections or immunocompromised patients. Fluconazole remains the most commonly prescribed systemic azole because of its favorable pharmacokinetics and oral bioavailability (Rex, et al., 1994).

Itraconazole, voriconazole, posaconazole, and echinocandins may be required in refractory or resistant infections (Wiederhold, 2017). Amphotericin B remains an important option in severe systemic candidiasis but is associated with significant toxicity concerns.

Successful management additionally requires elimination of predisposing factors whenever possible. Optimization of glycemic control, denture hygiene improvement, smoking cessation, xerostomia management, and reduction of unnecessary antibiotic exposure are critically important supportive measures (Soysa, Samaranayake & Ellepola, 2006).

Despite therapeutic effectiveness, recurrence remains common in immunocompromised populations. Persistent immunosuppression, biofilm formation, and antifungal resistance significantly complicate long-term disease control (de Repentigny, Lewandowski & Jolicoeur, 2004).

Antifungal Resistance and Biofilm-Associated Challenges

Antifungal resistance has emerged as one of the most important contemporary challenges in oral candidiasis management. Repeated antifungal exposure, prolonged azole therapy, and increasing prevalence of non-albicans *Candida* species have significantly contributed to resistance development (Perlin, 2015).

Azole resistance is particularly problematic because fluconazole remains the most commonly used systemic antifungal agent. Resistance mechanisms include efflux pump overexpression, alterations in ergosterol biosynthesis pathways, target enzyme mutations, and stress-response activation (Sanglard & Odds, 2002).

Candida glabrata demonstrates particular clinical importance because of its reduced susceptibility to azole antifungals and ability

to rapidly acquire multidrug resistance (Whaley & Rogers, 2016). Similarly, *Candida krusei* possesses intrinsic fluconazole resistance.

Biofilm-associated infections represent another major therapeutic challenge. *Candida* biofilms demonstrate dramatically increased resistance compared with planktonic cells, often requiring antifungal concentrations hundreds of times higher for eradication (Chandra, et al., 2001).

The extracellular biofilm matrix acts as a physical barrier that limits antifungal penetration. Additionally, biofilm-associated cells exhibit altered metabolic states and stress-response mechanisms that further enhance survival (Gulati & Nobile, 2016).

Mixed bacterial-fungal biofilms may demonstrate even greater resistance potential. Interactions between *Candida* species and oral bacteria may enhance microbial adhesion, virulence, and antifungal tolerance (Harriott & Noverr, 2009).

Denture-associated candidiasis exemplifies the clinical importance of biofilm-mediated infection. Acrylic denture surfaces provide favorable environments for persistent *Candida* colonization and recurrent infection (Pereira-Cenci, et al., 2008).

Because of increasing resistance concerns, routine species identification and susceptibility testing are becoming increasingly important in recurrent or refractory cases. Novel therapeutic strategies specifically targeting biofilms and resistant fungal strains are therefore urgently needed.

Novel and Emerging Therapeutic Approaches

The limitations of conventional antifungal therapies have stimulated substantial research into novel treatment strategies for oral candidiasis. Emerging approaches focus not only on fungal eradication but also on biofilm disruption, immune modulation, and restoration of microbial balance (Vila, et al., 2020).

Photodynamic therapy (PDT) has gained considerable attention as a promising antifungal modality. PDT involves activation of photosensitizing agents by specific light wavelengths, generating reactive oxygen species capable of destroying fungal cells (Mima, et al., 2012). Studies have demonstrated significant antifungal and antibiofilm effects against *Candida* species.

Photobiomodulation has additionally been investigated because of its wound healing and anti-inflammatory properties. Although PBM does not directly eradicate fungal organisms to the same extent as PDT, it may improve mucosal healing and reduce inflammation associated with candidiasis (de Senna, et al., 2018).

Nanotechnology-based antifungal systems represent another rapidly developing field. Nanoparticles may improve antifungal drug delivery, enhance mucosal penetration, and increase biofilm disruption capacity (Monteiro, et al., 2011). Silver nanoparticles, chitosan nanoparticles, and liposomal drug delivery systems have demonstrated promising antifungal activity.

Natural compounds and phytotherapeutic agents have also gained increasing interest. Essential oils, curcumin, propolis, tea tree oil, and thymoquinone have shown antifungal and antibiofilm effects in experimental studies (Shrestha & Kishen, 2014). Combination therapies involving natural compounds and conventional antifungals may additionally reduce resistance development.

Probiotic therapy represents another important emerging strategy. Beneficial bacterial strains such as *Lactobacillus* species may inhibit *Candida* colonization through competitive interactions and antimicrobial metabolite production (Hatakka, et al., 2007). Probiotics may help restore oral microbial balance and reduce recurrence risk.

Antimicrobial peptides are increasingly being investigated because of their broad-spectrum antifungal properties and reduced

resistance potential (Tsai, et al., 2011). Salivary histatins and synthetic peptide analogs may become valuable therapeutic agents in the future.

Immunotherapy and host-response modulation also represent promising future directions. Enhancement of mucosal immunity and cytokine regulation may improve host resistance against recurrent *Candida* infection (Conti & Gaffen, 2010).

Artificial intelligence-assisted antifungal drug discovery and fungal genomics are additionally accelerating development of targeted antifungal therapies (Perfect, 2012). Personalized antifungal treatment strategies based on species identification, resistance profiling, and host immune status may become increasingly important in the future.

Prevention Strategies in Immunocompromised Patients

Prevention of oral candidiasis is critically important in immunocompromised populations because recurrent infections may significantly impair quality of life and increase systemic infection risk. Preventive approaches should focus on reduction of fungal colonization, optimization of oral hygiene, maintenance of mucosal integrity, and control of underlying predisposing factors (Patil, et al., 2015).

Oral hygiene maintenance remains one of the most fundamental preventive strategies. Regular tooth brushing, tongue cleaning, professional dental care, and denture hygiene are essential for reducing biofilm accumulation and fungal colonization (Budtz-Jorgensen, 2000). Dentures should be cleaned thoroughly and preferably removed during nighttime to reduce continuous mucosal exposure to fungal biofilms.

Management of xerostomia is particularly important because reduced salivary flow significantly increases *Candida* colonization

risk. Saliva substitutes, salivary stimulants, hydration, and avoidance of dehydrating medications when possible may improve oral defense mechanisms (Sreebny & Schwartz, 1997).

Inhaled corticosteroid users should rinse their mouths after medication use to minimize local immunosuppressive effects and fungal overgrowth (Toogood, 1990). Similarly, optimization of glycemic control is essential in diabetic patients because hyperglycemia promotes *Candida* proliferation and impairs neutrophil function.

Nutritional support also plays a major role in maintaining mucosal immunity. Malnutrition, vitamin deficiencies, and iron deficiency anemia may increase susceptibility to oral fungal infections (Paillaud, et al., 2004).

Cancer patients receiving chemotherapy or radiotherapy may benefit from preventive oral care protocols including chlorhexidine rinses, low-level laser therapy for mucositis prevention, and regular oral examination (Sonis & Fey, 2002). Early identification of fungal colonization may help prevent progression to severe infection.

Prophylactic antifungal therapy is sometimes used in high-risk populations such as hematopoietic stem cell transplant recipients and severely immunocompromised oncology patients (Wingard, 1995). However, prolonged prophylactic antifungal use may contribute to resistance development and should therefore be carefully balanced.

Microbiome-preserving strategies are increasingly recognized as important preventive measures. Avoidance of unnecessary broad-spectrum antibiotic use may help maintain bacterial-fungal balance within the oral cavity (Pappas, 2006).

Education of patients and caregivers remains essential for successful prevention. Awareness regarding oral hygiene, denture

maintenance, symptom recognition, and risk factor modification may substantially improve long-term disease control.

Future Perspectives

The future management of oral candidiasis in immunocompromised patients will likely become increasingly personalized, multidisciplinary, and technology-driven. Rapid advances in molecular diagnostics, microbiome analysis, artificial intelligence, nanotechnology, and immunotherapy are expected to significantly improve both diagnosis and treatment (Bongomin, et al., 2017).

Precision medicine approaches may allow individualized antifungal therapy based on fungal species identification, resistance gene analysis, host immune status, and microbiome composition (Arendrup, 2014). Such strategies could improve therapeutic success while reducing unnecessary antifungal exposure and resistance development.

Artificial intelligence-assisted diagnostics may further enhance early detection and treatment planning. Machine learning systems analyzing oral images, salivary biomarkers, and microbiological profiles may facilitate rapid chairside diagnosis and risk prediction (Esteva, et al., 2019).

Nanotechnology-based drug delivery systems are expected to improve antifungal bioavailability, mucosal penetration, and sustained release characteristics (Gharat, Momin & Bhavsar, 2021). Targeted nanoparticle systems may additionally reduce systemic toxicity and improve efficacy against resistant biofilms.

Biomaterial science and tissue engineering may contribute to development of antifungal dentures, implant coatings, and mucosal protective materials capable of inhibiting fungal adhesion and biofilm formation (Song, et al., 2005).

Immunomodulatory therapies are likely to gain increasing importance in recurrent and refractory candidiasis. Enhancement of host mucosal immunity and targeted cytokine modulation may provide alternative therapeutic strategies beyond direct fungal eradication (Richardson, et al., 2019).

Microbiome-targeted therapies including probiotics, prebiotics, and microbial transplantation approaches may also emerge as important preventive and therapeutic modalities (Devine & Marsh, 2009). Restoration of balanced oral microbial ecology may reduce *Candida* pathogenicity and recurrence rates.

Development of novel antifungal agents remains critically important because resistance continues to increase globally. New drug classes targeting fungal virulence factors, quorum sensing pathways, and biofilm-associated mechanisms may significantly improve future treatment outcomes (Robbins, Caplan & Cowen, 2017).

Continued interdisciplinary collaboration among oral surgeons, microbiologists, immunologists, infectious disease specialists, and biomedical researchers will be essential for successful translation of emerging therapies into clinical practice.

Conclusion

Oral candidiasis remains one of the most significant opportunistic infections affecting immunocompromised patients and continues to present substantial diagnostic and therapeutic challenges. Increasing prevalence of immunosuppressive conditions, aging populations, widespread antimicrobial exposure, and emergence of resistant *Candida* species have contributed to growing clinical importance of fungal oral diseases.

Although *Candida albicans* remains the predominant pathogen, non-*albicans* *Candida* species such as *Candida glabrata*

and *Candida krusei* are becoming increasingly important because of their antifungal resistance profiles and biofilm-forming capabilities. Biofilm-associated infections represent a particularly difficult therapeutic problem because of enhanced microbial resistance and recurrent disease potential.

The pathogenesis of oral candidiasis involves highly complex interactions among fungal virulence factors, host immune dysfunction, salivary alterations, microbiome imbalance, and local environmental conditions. Understanding these multifactorial mechanisms is essential for development of more effective preventive and therapeutic strategies.

Conventional antifungal therapies including nystatin, azoles, and polyenes remain fundamental treatment modalities; however, recurrence and resistance continue to limit long-term success in many immunocompromised patients. Consequently, substantial research has focused on novel therapeutic approaches including photodynamic therapy, photobiomodulation, nanotechnology-based systems, probiotics, natural compounds, antimicrobial peptides, and immunomodulatory therapies.

Advances in molecular diagnostics, salivary biomarkers, fungal genomics, and artificial intelligence are expected to significantly improve future diagnostic precision and personalized treatment planning. Emerging regenerative and microbiome-targeted approaches may additionally transform long-term disease prevention and management.

Overall, successful management of oral candidiasis in immunocompromised patients requires comprehensive multidisciplinary care involving accurate diagnosis, risk factor control, optimization of host immunity, effective antifungal therapy, and long-term preventive strategies. Continued translational research and technological innovation will remain essential for

overcoming the growing challenges associated with fungal resistance and recurrent infection.

References

Akpan, A., & Morgan, R. (2002). Oral candidiasis. *Postgraduate Medical Journal*, 78(922), 455–459.

Arendorf, T. M., & Walker, D. M. (1980). Oral candidal populations in health and disease. *British Dental Journal*, 147(10), 267–272.

Arendrup, M. C. (2014). *Candida* and candidiasis susceptibility and epidemiology. *Clinical Microbiology and Infection*, 20(Suppl. 6), 42–48.

Bongomin, F., et al. (2017). Global and multi-national prevalence of fungal diseases. *Journal of Fungi*, 3(4), 57.

Brown, G. D., et al. (2012). Hidden killers: Human fungal infections. *Science Translational Medicine*, 4(165), 165rv13.

Budtz-Jorgensen, E. (1990). Etiology, pathogenesis, therapy, and prophylaxis of oral yeast infections. *Acta Odontologica Scandinavica*, 48(1), 61–69.

Budtz-Jorgensen, E. (2000). Ecology of *Candida*-associated denture stomatitis. *Microbial Ecology in Health and Disease*, 12(3), 170–185.

Calderone, R. A. (2002). *Candida and candidiasis*. ASM Press.

Calderone, R. A., & Clancy, C. J. (2012). *Candida and candidiasis* (2nd ed.). ASM Press.

Chandra, J., et al. (2001). Antifungal resistance of candidal biofilms. *Antimicrobial Agents and Chemotherapy*, 45(4), 1170–1175.

Coco, B. J., et al. (2008). Mixed *Candida albicans* and *Candida glabrata* populations associated with the pathogenesis of

denture stomatitis. *Oral Microbiology and Immunology*, 23(5), 377–383.

Conti, H. R., & Gaffen, S. L. (2010). IL-17-mediated immunity to oral candidiasis. *Journal of Dental Research*, 89(11), 1158–1169.

Coronado-Castellote, L., & Jiménez-Soriano, Y. (2013). Clinical and microbiological diagnosis of oral candidiasis. *Journal of Clinical and Experimental Dentistry*, 5(5), e279–e286.

de Repentigny, L., Lewandowski, D., & Jolicoeur, P. (2004). Immunopathogenesis of oropharyngeal candidiasis in HIV infection. *Clinical Microbiology Reviews*, 17(4), 729–759.

de Senna, A. M., et al. (2018). Photobiomodulation therapy in oral candidiasis. *Lasers in Medical Science*, 33(5), 1145–1152.

Devine, D. A., & Marsh, P. D. (2009). Prospects for the development of probiotics and prebiotics for oral applications. *Journal of Oral Microbiology*, 1(1), 1949.

Edgerton, M., & Koshlukova, S. E. (2000). Salivary histatins and host defense against *Candida*. *Current Opinion in Infectious Diseases*, 13(6), 643–648.

Ellepola, A. N., & Samaranayake, L. P. (2000). Oral candidal infections and antimycotics. *Critical Reviews in Oral Biology & Medicine*, 11(2), 172–198.

Epstein, J. B., & Polsky, B. (1998). Oropharyngeal candidiasis: A review of its clinical spectrum and current therapies. *Clinical Therapeutics*, 20(1), 40–57.

Esteva, A., et al. (2019). A guide to deep learning in healthcare. *Nature Medicine*, 25(1), 24–29.

Eversole, L. R. (2001). *Clinical outline of oral pathology* (4th ed.). PMPH-USA.

Farah, C. S., Lynch, N., & McCullough, M. J. (2010). Oral fungal infections: An update for the general practitioner. *Australian Dental Journal*, 55(Suppl. 1), 48–54.

Fidel, P. L., Jr., Vazquez, J. A., & Sobel, J. D. (1999). *Candida glabrata*: Review of epidemiology, pathogenesis and clinical disease. *Clinical Microbiology Reviews*, 12(1), 80–96.

Fox, R. I. (2005). Sjögren syndrome. *The Lancet*, 366(9482), 321–331.

Gharat, S. A., Momin, M., & Bhavsar, C. (2021). Oral candidiasis and novel drug delivery systems. *Journal of Drug Delivery Science and Technology*, 63, 102559.

Gladiator, A., et al. (2013). Cutting edge: IL-17-secreting innate lymphoid cells are essential for host defense against fungal infection. *The Journal of Immunology*, 190(2), 521–525.

Gonsalves, W. C., Chi, A. C., & Neville, B. W. (2007). Common oral lesions: Part I. Superficial mucosal lesions. *American Family Physician*, 75(4), 501–507.

Greenspan, D. (1997). Oral manifestations of HIV infection. *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology*, 84(1), 102–110.

Guggenheimer, J., et al. (2000). Insulin-dependent diabetes mellitus and oral soft tissue pathologies. *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology*, 89(5), 563–569.

Gulati, M., & Nobile, C. J. (2016). *Candida albicans* biofilms: Development, regulation, and molecular mechanisms. *Microbes and Infection*, 18(5), 310–321.

Harriott, M. M., & Noverr, M. C. (2009). *Candida albicans* and *Staphylococcus aureus* form polymicrobial biofilms. *Infection and Immunity*, 77(9), 3914–3927.

Hatakka, K., et al. (2007). Probiotics reduce prevalence of oral *Candida* in elderly. *Journal of Dental Research*, 86(2), 125–130.

Hube, B. (2004). From commensal to pathogen: Stage- and tissue-specific gene expression of *Candida albicans*. *Current Opinion in Microbiology*, 7(4), 336–341.

Imanguli, M. M., et al. (2008). Oral graft-versus-host disease. *Oral Diseases*, 14(5), 396–412.

Jenkinson, H. F., & Lamont, R. J. (2005). Oral microbial communities in sickness and in health. *Trends in Microbiology*, 13(12), 589–595.

Kanbe, T., et al. (2002). PCR-based identification of medically important *Candida* species. *Journal of Clinical Microbiology*, 40(7), 2399–2403.

Lyu, X., et al. (2016). Efficacy of nystatin for the treatment of oral candidiasis. *Drug Design, Development and Therapy*, 10, 1161–1171.

Mayer, F. L., Wilson, D., & Hube, B. (2013). *Candida albicans* pathogenicity mechanisms. *Virulence*, 4(2), 119–128.

Mima, E. G., et al. (2012). Comparison of photodynamic therapy versus conventional antifungal therapy for denture stomatitis. *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology*, 114(4), 469–476.

Monteiro, D. R., et al. (2011). Silver nanoparticles: Influence of stabilizing agent and diameter on antifungal activity. *International Journal of Antimicrobial Agents*, 38(6), 523–527.

Naglik, J. R., Challacombe, S. J., & Hube, B. (2003). *Candida albicans* secreted aspartyl proteinases in virulence and pathogenesis. *Microbiology and Molecular Biology Reviews*, 67(3), 400–428.

Netea, M. G., et al. (2015). Immune defence against *Candida* fungal infections. *Nature Reviews Immunology*, 15(10), 630–642.

Nett, J. E., & Andes, D. R. (2020). Contributions of the biofilm matrix to *Candida* pathogenesis. *Journal of Fungi*, 6(1), 21.

Neville, B. W., et al. (2016). *Oral and maxillofacial pathology* (4th ed.). Elsevier.

Odds, F. C. (1988). *Candida and candidosis* (2nd ed.). Baillière Tindall.

Paillaud, E., et al. (2004). Oral candidiasis and nutritional deficiencies in elderly hospitalized patients. *British Journal of Nutrition*, 92(5), 861–867.

Pappas, P. G. (2006). Invasive candidiasis. *Infectious Disease Clinics of North America*, 20(3), 485–506.

Pappas, P. G., et al. (2016). Clinical practice guideline for the management of candidiasis. *Clinical Infectious Diseases*, 62(4), e1–e50.

Patil, S., et al. (2015). Clinical appearance of oral *Candida* infection and therapeutic strategies. *Frontiers in Microbiology*, 6, 1391.

Pereira-Cenci, T., et al. (2008). Development of *Candida*-associated denture stomatitis. *Journal of Applied Oral Science*, 16(2), 86–94.

Perfect, J. R. (2012). The impact of the host on fungal infections. *The American Journal of Medicine*, 125(1 Suppl), S39–S51.

Perfect, J. R. (2017). The antifungal pipeline: A reality check. *Nature Reviews Drug Discovery*, 16(9), 603–616.

Perlin, D. S. (2015). Mechanisms of echinocandin antifungal drug resistance. *Annals of the New York Academy of Sciences*, 1354(1), 1–11.

Pfaller, M. A., & Diekema, D. J. (2007). Epidemiology of invasive candidiasis. *Clinical Microbiology Reviews*, 20(1), 133–163.

Pien, F. D., et al. (1991). Fluconazole treatment of candidiasis. *Mayo Clinic Proceedings*, 66(10), 1018–1024.

Ramage, G., et al. (2004). Denture stomatitis: A role for *Candida* biofilms. *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology*, 98(1), 53–59.

Ramage, G., Martínez, J. P., & López-Ribot, J. L. (2006). *Candida* biofilms on implanted biomaterials. *Clinical Microbiology Reviews*, 19(1), 167–183.

Reamy, B. V., Derby, R., & Bunt, C. W. (2010). Common tongue conditions in primary care. *American Family Physician*, 81(5), 627–634.

Rex, J. H., et al. (1994). A randomized trial comparing fluconazole with amphotericin B for candidemia. *The New England Journal of Medicine*, 331(20), 1325–1330.

Richardson, J. P., et al. (2019). *Candida* innate immunity at the mucosa. *Seminars in Cell & Developmental Biology*, 89, 58–70.

Robbins, N., Caplan, T., & Cowen, L. E. (2017). Molecular evolution of antifungal drug resistance. *Annual Review of Microbiology*, 71, 753–775.

Samaranayake, L. P. (1992). Oral mycoses in HIV infection. *Oral Surgery, Oral Medicine, Oral Pathology*, 73(2), 171–180.

Samaranayake, L. P., & Holmstrup, P. (1989). Oral candidiasis and human immunodeficiency virus infection. *Journal of Oral Pathology & Medicine*, 18(10), 554–564.

Sanglard, D., & Odds, F. C. (2002). Resistance of *Candida* species to antifungal agents. *The Lancet Infectious Diseases*, 2(2), 73–85.

Schwendicke, F., et al. (2019). Convolutional neural networks for dental image diagnostics. *Journal of Dental Research*, 98(13), 1465–1471.

Sharon, V., & Fazel, N. (2010). Oral candidiasis and angular cheilitis. *Dermatologic Therapy*, 23(3), 230–242.

Shrestha, A., & Kishen, A. (2014). Antibiofilm efficacy of photosensitizer-functionalized bioactive nanoparticles on multispecies biofilm. *Journal of Endodontics*, 40(10), 1604–1610.

Silva, S., et al. (2012). *Candida glabrata*, *Candida parapsilosis* and *Candida tropicalis*: Biology, epidemiology and pathogenicity. *FEMS Microbiology Reviews*, 36(2), 288–305.

Sitheeque, M. A. M., & Samaranayake, L. P. (2003). Chronic hyperplastic candidosis/candidiasis. *Critical Reviews in Oral Biology & Medicine*, 14(4), 253–267.

Song, X., et al. (2005). Genetic relatedness of oral yeasts within and between patients with marginal periodontitis and periodontally healthy subjects. *Oral Microbiology and Immunology*, 20(1), 25–31.

Sonis, S. T. (2004). Oral mucositis in cancer therapy. *Journal of Supportive Oncology*, 2(6 Suppl. 3), 3–8.

Sonis, S. T., & Fey, E. G. (2002). Oral complications of cancer therapy. *Oncology*, 16(5 Suppl. 4), 680–695.

Soysa, N. S., & Ellepola, A. N. (2005). The impact of cigarette/tobacco smoking on oral candidosis. *Oral Diseases*, 11(5), 268–273.

Soysa, N. S., Samaranayake, L. P., & Ellepola, A. N. B. (2006). Diabetes mellitus as a contributory factor in oral candidosis. *Diabetic Medicine*, 23(5), 455–459.

Sreebny, L. M., & Schwartz, S. S. (1997). A reference guide to drugs and dry mouth. *Gerodontology*, 14(1), 33–47.

Sudbery, P., Gow, N., & Berman, J. (2004). The distinct morphogenic states of *Candida albicans*. *Trends in Microbiology*, 12(7), 317–324.

Sundstrom, P. (2002). Adhesion in *Candida* spp. *Cellular Microbiology*, 4(8), 461–469.

Toogood, J. H. (1990). Complications of topical steroid therapy for asthma. *American Review of Respiratory Disease*, 141(2), S89–S96.

Tsai, P. W., et al. (2011). Human antimicrobial peptide LL-37 inhibits adhesion of *Candida albicans*. *Antimicrobial Agents and Chemotherapy*, 55(7), 3511–3519.

Vila, T., et al. (2020). Oral candidiasis: A disease of opportunity. *Journal of Fungi*, 6(1), 15.

Webb, B. C., et al. (1998). *Candida*-associated denture stomatitis. *Oral Microbiology and Immunology*, 13(3), 130–139.

Whaley, S. G., & Rogers, P. D. (2016). Azole resistance in *Candida glabrata*. *Current Infectious Disease Reports*, 18(12), 41.

Wiederhold, N. P. (2017). Antifungal resistance: Current trends and future strategies. *Infection and Drug Resistance*, 10, 249–259.

Williams, D. W., & Lewis, M. A. O. (2000). Oral microbiology: Isolation and identification of *Candida* from the oral cavity. *Oral Diseases*, 6(1), 3–11.

Williams, D., & Lewis, M. (2011). Pathogenesis and treatment of oral candidosis. *Journal of Oral Microbiology*, 3(1), 5771.

Wingard, J. R. (1995). Importance of *Candida* species other than *Candida albicans* as pathogens in oncology patients. *Clinical Infectious Diseases*, 20(1), 115–125.

Xu, H., Jenkinson, H. F., & Dongari-Bagtzoglou, A. (2014). Innocent until proven guilty: Mechanisms and roles of *Streptococcus*-*Candida* interactions in oral health and disease. *Molecular Oral Microbiology*, 29(3), 99–116.

CHAPTER 2

3D Printing Technologies in Oral and Maxillofacial Surgery

Özge ÖZDAL ZİNCİR¹

Introduction

Three-dimensional (3D) printing technologies have revolutionized contemporary oral and maxillofacial surgery by enabling highly precise, patient-specific, and digitally driven treatment approaches. The integration of advanced imaging modalities, computer-aided design/computer-aided manufacturing (CAD/CAM) systems, virtual surgical planning, and additive manufacturing has significantly transformed surgical workflows in maxillofacial reconstruction, implantology, trauma surgery, orthognathic surgery, and regenerative medicine (Martelli, et al. 2016).

Traditionally, oral and maxillofacial surgical procedures relied heavily on two-dimensional imaging, manual model fabrication, and intraoperative adaptation of fixation devices. Although these conventional methods remain effective, they may be associated with increased surgical time, limited precision, and

¹ Assoc. Prof. Dr. Istanbul University- Cerrahpasa, Faculty of Dentistry, Department of Oral and Maxillofacial Surgery ORCID: 0009-0000-9423-5764

unpredictable outcomes in complex anatomical situations (Winder & Bibb, 2005). The emergence of 3D printing technologies has therefore provided clinicians with new opportunities for personalized surgical planning and improved anatomical accuracy.

Three-dimensional printing, also known as additive manufacturing, refers to the layer-by-layer fabrication of physical objects based on digital models (Hull, 1986). In oral and maxillofacial surgery, digital data obtained from computed tomography (CT), cone-beam computed tomography (CBCT), magnetic resonance imaging (MRI), or intraoral scanning may be converted into printable models and patient-specific devices (Dawood, et al., 2015). Such technologies allow surgeons to fabricate surgical guides, anatomical models, customized implants, reconstruction plates, prosthetic frameworks, tissue scaffolds, and bioprinted constructs.

One of the most important advantages of 3D printing involves patient-specific customization. Because maxillofacial anatomy demonstrates substantial individual variation, personalized surgical devices may significantly improve functional and esthetic outcomes (Mazzoni, et al., 2015). Patient-specific implants and reconstruction plates may reduce intraoperative plate bending, improve anatomical adaptation, shorten surgical time, and enhance surgical precision.

Another major application involves surgical simulation and education. Anatomical models generated through 3D printing allow surgeons to rehearse complex procedures preoperatively and facilitate communication among surgical teams, patients, and trainees (Choi & Kim, 2015). These models have become increasingly valuable in resident education and surgical training programs.

The field of bioprinting has additionally emerged as one of the most promising frontiers in regenerative medicine. Unlike conventional 3D printing, bioprinting utilizes bioinks containing living cells, biomaterials, and growth factors to fabricate biologically functional tissues (Murphy & Atala, 2014). In oral and maxillofacial surgery, bioprinting may eventually enable fabrication of bone, cartilage, periodontal tissues, and even vascularized tissue constructs.

Despite remarkable technological progress, important challenges remain regarding cost, material limitations, printing accuracy, regulatory approval, long-term clinical outcomes, and biological integration (Javaid & Haleem, 2019). Nevertheless, continuous innovation in biomaterials, software technologies, artificial intelligence integration, and tissue engineering is expected to further expand the role of 3D printing in oral and maxillofacial surgery.

This chapter reviews the principles, technologies, clinical applications, advantages, limitations, and future perspectives of 3D printing technologies in oral and maxillofacial surgery.

Principles of 3D Printing Technologies

Three-dimensional printing is based on additive manufacturing principles in which physical objects are created through sequential layer deposition according to digital design data (Gibson, Rosen & Stucker, 2015). The process generally begins with acquisition of anatomical imaging data obtained from CT, CBCT, MRI, or optical scanning systems. These datasets are converted into digital 3D models using specialized segmentation software.

After digital segmentation, computer-aided design software is used to modify and optimize the virtual model according to clinical requirements (Mangano, et al., 2017). The finalized digital model is

then exported in stereolithography (STL) format, which serves as the standard file type for most 3D printing systems.

Several additive manufacturing technologies are currently used in oral and maxillofacial surgery. Stereolithography (SLA) is among the earliest and most widely utilized techniques. SLA printers use ultraviolet laser energy to polymerize liquid photopolymer resins layer by layer (Alharbi, Osman & Wismeijer, 2016). This technology provides excellent surface accuracy and is commonly used for surgical guides, anatomical models, and dental applications.

Fused deposition modeling (FDM) represents another commonly used method. FDM printers fabricate objects through extrusion of thermoplastic materials such as polylactic acid or acrylonitrile butadiene styrene (Hazeveld, Huddleston Slater, & Ren, 2014). Although relatively affordable, FDM systems may demonstrate lower printing resolution compared with SLA systems.

Selective laser sintering (SLS) and selective laser melting (SLM) are advanced powder-based technologies capable of producing highly durable metallic structures (Van Noort, 2012). These systems are especially important for fabrication of titanium patient-specific implants and reconstruction plates.

Electron beam melting (EBM) is another metal printing technology frequently used for biomedical implants. EBM allows fabrication of porous titanium structures with excellent mechanical properties and osseointegration potential (Murr, et al., 2010).

Material jetting and binder jetting technologies are additionally used for high-resolution printing applications and multicomponent fabrication (Gross, et al., 2014). Continuous improvements in printer resolution, material compatibility, and printing speed continue to expand the clinical potential of additive manufacturing systems.

Material selection represents a critical aspect of 3D printing in surgery. Commonly used materials include photopolymer resins, thermoplastics, ceramics, titanium alloys, cobalt-chromium alloys, and biodegradable biomaterials (Javaid & Haleem, 2018). In regenerative applications, bioactive scaffolds and hydrogel-based bioinks have become increasingly important.

The accuracy of 3D printed structures depends on multiple factors including image quality, segmentation precision, software processing, printer calibration, layer thickness, and material properties (Tahayeri, et al., 2018). Consequently, strict quality control protocols remain essential in clinical applications.

Virtual Surgical Planning and Digital Workflow

Virtual surgical planning (VSP) has become one of the most important components of modern digital oral and maxillofacial surgery. VSP combines radiological imaging, computer-assisted design, and additive manufacturing to optimize surgical precision and predictability (Xia, Gateno & Teichgraber, 2009).

The workflow generally begins with acquisition of high-resolution imaging data. CT and CBCT scans are commonly used because of their ability to provide detailed anatomical visualization of hard tissues (Swennen, Mollemans & Schutyser, 2009). In some cases, intraoral scanning and facial scanning may additionally be integrated to improve soft tissue and dental accuracy.

After image acquisition, digital segmentation is performed to isolate relevant anatomical structures such as bone, teeth, tumors, or neurovascular components (Bell, 2010). Specialized software allows surgeons to manipulate virtual anatomical models, simulate osteotomies, evaluate reconstruction options, and fabricate customized surgical devices.

Virtual planning is particularly valuable in orthognathic surgery and maxillofacial reconstruction. Surgical movements may be simulated digitally, allowing more accurate prediction of postoperative skeletal relationships and facial symmetry (Zinser, et al., 2014). Surgical splints generated through 3D printing may subsequently transfer the virtual plan accurately to the operative field.

One of the major advantages of digital workflow integration is reduction of intraoperative uncertainty. Surgeons may rehearse procedures preoperatively, identify anatomical limitations, and optimize reconstruction strategies before surgery (Modabber, et al., 2012).

Communication among multidisciplinary teams may also improve substantially through virtual planning. Surgeons, prosthodontists, radiologists, biomedical engineers, and laboratory technicians may collaboratively evaluate digital models and treatment options (Ciocca, et al., 2010).

Artificial intelligence integration has additionally begun influencing digital workflows. AI-assisted segmentation and automated surgical planning algorithms may further improve efficiency and reduce operator dependency in the future (Schwendicke, Samek & Krois, 2020).

3D Printed Surgical Guides

Surgical guides represent one of the most widely utilized clinical applications of 3D printing in oral surgery and implantology. These guides are fabricated according to virtual surgical plans and are designed to transfer digital planning accurately into the clinical setting (Vercruyssen, et al., 2014).

In implant dentistry, static surgical guides allow precise implant positioning according to prosthetically driven treatment

plans. Guided implant surgery may improve implant angulation, depth control, and anatomical safety while reducing surgical invasiveness (D'haese, et al., 2017).

Guided implant surgery is particularly advantageous in anatomically complex situations involving limited bone volume, proximity to vital structures, or full-arch rehabilitation (Tahmaseb, et al., 2014). Surgical guides may additionally facilitate flapless implant placement, thereby reducing postoperative morbidity and accelerating healing.

Accuracy remains one of the most important considerations in guided surgery. Multiple factors including CBCT quality, software alignment, guide stability, sleeve design, and printing accuracy may influence surgical precision (Schneider, et al., 2009). Nevertheless, numerous studies have demonstrated improved implant placement accuracy compared with freehand techniques.

Beyond implantology, surgical guides are increasingly utilized in orthognathic surgery, reconstructive surgery, and tumor resection (Metzger, et al., 2007). Osteotomy guides may improve surgical precision and reduce operative time in mandibular and maxillary osteotomies.

Tumor surgery represents another important indication. Patient-specific cutting guides may facilitate accurate tumor margin resection while preserving critical anatomical structures (Zweifel, et al., 2015). Such approaches are particularly valuable in complex mandibular reconstruction procedures.

Endodontic microsurgery and autotransplantation procedures have also benefited from guided surgical technologies. Three-dimensional printed guides may improve access precision and reduce procedural complications (Strbac, et al., 2017).

Despite significant advantages, guided surgery systems may increase preoperative preparation time and cost. Additionally, inaccuracies may still occur because of guide movement, manufacturing errors, or limited mouth opening during surgery (Van Assche, et al., 2012). Continued refinement of digital workflows is therefore necessary to improve reliability and accessibility.

Patient-Specific Implants in Oral and Maxillofacial Surgery

Patient-specific implants (PSIs) represent one of the most transformative applications of 3D printing technologies in oral and maxillofacial surgery. Unlike conventional stock implants that require extensive intraoperative adaptation, PSIs are individually designed according to the patient's anatomical characteristics and surgical requirements (Zweifel, et al., 2014).

The development of PSIs typically begins with acquisition of high-resolution CT or CBCT data followed by digital segmentation and virtual reconstruction. In unilateral defects, mirroring techniques may be used to reproduce the unaffected anatomical side and create symmetrical reconstructions (Modabber, et al., 2012). CAD software subsequently allows fabrication of customized implants precisely matching the planned anatomy.

Titanium remains the most widely used material for maxillofacial PSIs because of its excellent biocompatibility, corrosion resistance, mechanical strength, and osseointegration capacity (Jardini, et al., 2014). Additive manufacturing techniques such as selective laser melting and electron beam melting enable fabrication of highly complex titanium structures with porous architectures and individualized geometries.

Patient-specific implants have become increasingly important in mandibular and midfacial reconstruction following trauma, tumor resection, congenital deformities, and severe atrophy (Wilde, et al., 2015). Customized implants may improve facial

symmetry, occlusal relationships, functional rehabilitation, and esthetic outcomes.

Mandibular reconstruction following oncological surgery is among the most important clinical indications for PSIs. Traditional freehand reconstruction using reconstruction plates may involve prolonged surgical time and less predictable anatomical adaptation (Roser, et al., 2010). In contrast, customized reconstruction systems may facilitate more accurate fibula flap positioning and improved mandibular contour restoration.

Orbital reconstruction represents another important application area. Orbital fractures and tumor-related orbital defects often require highly precise anatomical restoration to prevent enophthalmos, diplopia, and facial asymmetry (Kozakiewicz, et al., 2009). Patient-specific orbital implants may significantly improve reconstruction accuracy and orbital volume restoration.

Temporomandibular joint reconstruction has also benefited substantially from customized implant technologies. Patient-specific TMJ prostheses may improve joint stability, mandibular function, and long-term functional outcomes in severe joint pathology (Wolford, et al., 2015).

Another major advantage of PSIs is reduction of intraoperative plate bending and adaptation. Conventional reconstruction plates often require repeated manual adjustment during surgery, increasing operative time and potential metal fatigue (Toto, et al., 2015). Customized implants eliminate much of this intraoperative modification and may therefore improve surgical efficiency.

Despite these advantages, PSIs remain associated with important limitations including high manufacturing costs, longer preoperative planning periods, regulatory considerations, and limited intraoperative flexibility (Zweifel, et al., 2015). Additionally,

manufacturing inaccuracies or planning errors may significantly affect surgical outcomes because implants are fabricated specifically according to the preoperative virtual plan.

Long-term biomechanical behavior and stress distribution within customized implants also remain important research areas. Nevertheless, current evidence strongly supports the growing role of PSIs in personalized oral and maxillofacial surgery.

3D Printed Reconstruction Plates

Reconstruction plates are essential components in maxillofacial trauma surgery, tumor reconstruction, and corrective surgery. Conventional reconstruction plates typically require extensive intraoperative contouring to adapt to the patient's anatomy (Ellis & Graham, 2002). Such manual bending may increase operative time and produce inaccuracies in anatomical reconstruction.

Three-dimensional printing technologies have enabled the development of customized reconstruction plates designed specifically according to patient anatomy and surgical planning (Yang, et al., 2018). These plates may be fabricated directly through metal additive manufacturing or indirectly through preoperative plate bending on printed anatomical models.

Customized reconstruction plates are especially advantageous in mandibular continuity defects resulting from tumor resection or severe trauma. Precise anatomical adaptation may improve mandibular contour restoration, occlusal stability, and facial symmetry (Mazzoni, et al., 2013).

Pre-bent plates generated on 3D printed anatomical models may additionally reduce surgical duration and improve reconstruction accuracy. Surgeons may perform plate adaptation preoperatively rather than intraoperatively, thereby reducing

ischemia time during free flap reconstruction procedures (Foley, et al., 2010).

Biomechanical optimization represents another important advantage of digital plate design. Finite element analysis may be integrated into the digital workflow to optimize stress distribution and reduce implant failure risk (Zhang, et al., 2018).

Patient-specific reconstruction plates may additionally facilitate immediate dental rehabilitation through integration with implant planning and prosthetic workflows (Ciocca, et al., 2012). Such combined approaches are becoming increasingly important in functional jaw reconstruction.

Despite these advantages, mechanical complications including screw loosening, plate fracture, and stress concentration may still occur (Avraham, et al., 2014). Long-term clinical studies remain necessary to evaluate durability and complication rates of customized reconstruction systems.

Bioprinting and Tissue Engineering Applications

Bioprinting represents one of the most innovative and rapidly evolving areas in regenerative medicine and oral surgery. Unlike conventional 3D printing, bioprinting utilizes living cells, biomaterials, growth factors, and bioactive molecules to fabricate biologically functional tissue constructs (Murphy, De Coppi & Atala, 2020).

The ultimate goal of bioprinting is the creation of viable tissues capable of replacing damaged or missing anatomical structures. In oral and maxillofacial surgery, this technology may eventually allow regeneration of bone, cartilage, periodontal tissues, salivary glands, and vascularized soft tissues (Kang, et al., 2016).

Bioinks are among the most important components of bioprinting systems. These materials typically consist of hydrogels

combined with living cells and biological signaling molecules (Gungor-Ozkerim, et al., 2018). Commonly used bioinks include collagen, gelatin, alginate, fibrin, and hyaluronic acid-based materials.

Bone tissue engineering is one of the primary focuses of maxillofacial bioprinting research. Critical-size bone defects remain difficult to reconstruct using conventional grafting techniques because of donor site morbidity, limited graft availability, and unpredictable resorption (Oryan, et al., 2014). Bioprinted scaffolds may potentially overcome some of these limitations by providing customized osteoconductive and osteoinductive environments.

Mesenchymal stem cells are frequently incorporated into bioprinted constructs because of their osteogenic differentiation capacity (Pittenger, et al., 1999). Growth factors such as bone morphogenetic proteins and vascular endothelial growth factor may additionally enhance tissue regeneration and vascularization.

Vascularization remains one of the major challenges in bioprinting. Adequate blood supply is essential for survival of large tissue constructs (Kolesky, et al., 2014). Consequently, substantial research is currently focused on development of vascularized bioprinted tissues.

Cartilage bioprinting represents another promising field, particularly for temporomandibular joint reconstruction and craniofacial defects (Daly, et al., 2016). Customized cartilage constructs may eventually improve functional and esthetic outcomes in facial reconstructive surgery.

Periodontal tissue engineering has additionally gained increasing attention. Bioprinted periodontal scaffolds may potentially regenerate periodontal ligament, cementum, and alveolar bone simultaneously (Rasperini, et al., 2015). Such approaches could

significantly improve management of severe periodontal defects in the future.

Despite remarkable scientific progress, clinical application of bioprinting remains limited because of regulatory challenges, technical complexity, cost, and biological integration difficulties (Mandrycky, et al., 2016). Nevertheless, ongoing advances in stem cell biology, biomaterials, and tissue engineering continue to accelerate translational progress.

Educational and Training Applications of 3D Printing

Three-dimensional printing technologies have become increasingly valuable in oral and maxillofacial surgical education and training. Traditional educational approaches often rely on cadaveric dissection, plastic models, and textbook illustrations, each of which possesses important limitations (Rose, et al., 2015).

Three-dimensional printed anatomical models may provide highly accurate patient-specific representations of craniofacial anatomy and pathology. Such models allow residents and surgeons to visualize complex anatomical relationships more effectively than conventional two-dimensional imaging (McMenamin, et al., 2014).

Surgical simulation represents one of the most important educational applications. Complex procedures including orthognathic surgery, mandibular reconstruction, orbital reconstruction, and tumor resection may be rehearsed preoperatively using patient-specific printed models (George, et al., 2017). This may improve surgical confidence, reduce operative errors, and shorten learning curves.

Training in implant dentistry has also benefited substantially from 3D printed models. Printed models may replicate bone density variations, anatomical structures, and surgical scenarios, thereby

improving procedural training before patient treatment (Reymus, et al., 2019).

Another important advantage involves improved patient communication. Printed anatomical models may help patients better understand surgical procedures, pathology extent, and expected outcomes (Witowski, et al., 2018). Improved visualization may increase informed consent quality and patient satisfaction.

Three-dimensional printed educational models are additionally valuable in interdisciplinary communication among surgeons, prosthodontists, radiologists, and biomedical engineers (Tack, et al., 2016). Such collaborative planning is particularly important in complex reconstructive cases.

Cost-effectiveness and accessibility continue to improve as desktop printing technologies become more affordable. Consequently, educational applications of 3D printing are expected to expand significantly in oral and maxillofacial surgery residency programs worldwide (Lim, et al., 2016).

Limitations and Challenges of 3D Printing Technologies

Despite the substantial advantages of 3D printing technologies, several important limitations remain. One of the major challenges involves manufacturing cost. Advanced metal printing systems, software platforms, biomaterials, and maintenance requirements may represent significant financial investments (Javaid & Haleem, 2018).

Production time may also limit clinical efficiency in urgent surgical situations. Although printing speeds continue to improve, fabrication of complex patient-specific implants and anatomical models may still require several hours or days (Rengier, et al., 2010).

Printing accuracy is critically important because small deviations may significantly affect surgical outcomes. Errors may

occur during imaging acquisition, segmentation, software processing, printing, or sterilization procedures (Mitsouras, et al., 2015). Strict quality control protocols are therefore essential.

Material limitations remain another challenge. Although titanium and polymer-based materials are widely used, development of biomaterials capable of simultaneously providing optimal mechanical strength, biocompatibility, biodegradability, and regenerative potential remains ongoing (Bose, Vahabzadeh & Bandyopadhyay, 2013).

Regulatory and ethical considerations are increasingly important as personalized implants and bioprinted tissues become more clinically advanced. Manufacturing standards, sterilization protocols, long-term safety evaluation, and legal responsibility issues continue to evolve (Ventola, 2014).

In bioprinting applications, biological challenges including vascularization, immune response, cell survival, and long-term tissue integration remain substantial obstacles (Murphy & Atala, 2014). Translating experimental tissue engineering approaches into routine clinical practice will require further extensive research.

Operator expertise also plays a critical role in successful implementation. Digital workflow management, software manipulation, anatomical segmentation, and surgical planning require specialized training and interdisciplinary collaboration (Martelli, et al., 2016).

Despite these limitations, rapid technological progress continues to improve accessibility, precision, affordability, and clinical reliability of additive manufacturing systems.

Future Perspectives of 3D Printing in Oral and Maxillofacial Surgery

The future of 3D printing technologies in oral and maxillofacial surgery appears exceptionally promising and is expected to further transform personalized surgical care. Continuous advances in biomaterials, artificial intelligence, tissue engineering, robotics, and digital imaging are rapidly expanding the capabilities of additive manufacturing systems (Javaid & Haleem, 2019).

Artificial intelligence integration is likely to become one of the most influential developments in future digital workflows. AI-assisted segmentation, automated surgical planning, and predictive modeling may significantly reduce planning time and operator dependency while improving surgical precision (Litjens, et al., 2017). Machine learning systems may additionally help optimize implant design, stress distribution, and reconstruction strategies according to patient-specific anatomical and biomechanical factors.

Another major future direction involves fully integrated digital workflows combining intraoral scanning, facial scanning, CBCT imaging, CAD/CAM systems, navigation surgery, and robotic-assisted procedures (Eggbeer, et al., 2012). Such integrated systems may improve surgical predictability and allow real-time intraoperative guidance.

Bioprinting is expected to become increasingly important in regenerative oral surgery and craniofacial tissue engineering. Although current applications remain largely experimental, future developments may eventually enable fabrication of vascularized bone grafts, cartilage structures, periodontal tissues, and even complete biologically functional maxillofacial constructs (Grayson, et al., 2010).

Stem cell technologies and growth factor incorporation may significantly improve the regenerative capacity of bioprinted tissues.

Personalized regenerative constructs generated from autologous cells could potentially reduce immune rejection and improve long-term integration (Kaigler, et al., 2013).

Four-dimensional (4D) printing represents another emerging field. Unlike conventional static 3D printed structures, 4D printed materials may change shape or function over time in response to environmental stimuli such as temperature, moisture, or mechanical stress (Momeni, et al., 2017). Such smart biomaterials may eventually allow dynamic tissue adaptation and self-modifying implants.

Nanotechnology integration may additionally improve the biological performance of printed biomaterials. Nanostructured surfaces may enhance osteoblast attachment, antibacterial activity, angiogenesis, and osseointegration (Webster & Ejiogor, 2004).

Point-of-care manufacturing within hospital settings may also become increasingly common. In-house printing laboratories could potentially reduce production time, improve workflow efficiency, and increase accessibility of patient-specific devices (Giannopoulos, et al., 2016).

Educational applications are expected to expand substantially as virtual reality, augmented reality, and haptic simulation technologies become integrated with printed anatomical models (Barsom, Graafland & Schijven, 2016). Such multimodal educational systems may significantly enhance surgical training and preoperative simulation.

Despite the remarkable future potential, continued collaboration among surgeons, biomedical engineers, material scientists, regulatory authorities, and software developers will remain essential for successful clinical translation and standardization.

Conclusion

Three-dimensional printing technologies have fundamentally transformed modern oral and maxillofacial surgery by enabling personalized, digitally driven, and highly precise treatment approaches. The integration of advanced imaging, virtual surgical planning, CAD/CAM technologies, and additive manufacturing has significantly improved surgical accuracy, predictability, and patient-specific customization.

Among the most important clinical applications are surgical guides, patient-specific implants, customized reconstruction plates, anatomical models, and regenerative scaffolds. Guided surgery systems have enhanced implant placement precision and improved surgical safety, while customized implants and reconstruction plates have optimized anatomical reconstruction in trauma, oncological, and congenital deformity cases.

Three-dimensional printing has additionally become an invaluable educational and communication tool. Anatomical models facilitate surgical simulation, resident training, interdisciplinary planning, and patient education. Such applications contribute significantly to improved surgical understanding and procedural preparation.

Bioprinting represents one of the most exciting future directions in oral and maxillofacial surgery. Although still largely experimental, the potential to fabricate biologically functional tissues and vascularized constructs may eventually revolutionize regenerative craniofacial reconstruction.

Despite substantial advantages, important challenges remain regarding cost, printing accuracy, biomaterial limitations, production time, regulatory considerations, and long-term clinical validation. Standardization of digital workflows and continued technological refinement are therefore essential for broader clinical adoption.

Future developments involving artificial intelligence, nanotechnology, smart biomaterials, regenerative medicine, and integrated digital workflows are expected to further expand the role of additive manufacturing in oral surgery. As technologies continue to evolve, 3D printing is likely to become an increasingly central component of precision-based oral and maxillofacial surgical care.

References

Alharbi, N., Osman, R., & Wismeijer, D. (2016). Effects of build direction on the mechanical properties of 3D-printed complete coverage interim dental restorations. *The Journal of Prosthetic Dentistry*, 115(6), 760–767.

Avraham, T., et al. (2014). Functional outcomes of fibula free flap mandible reconstruction with patient-specific plates. *Journal of Oral and Maxillofacial Surgery*, 72(7), 1323–1331.

Barsom, E. Z., Graafland, M., & Schijven, M. P. (2016). Systematic review on the effectiveness of augmented reality applications in medical training. *Surgical Endoscopy*, 30(10), 4174–4183.

Bell, R. B. (2010). Computer planning and intraoperative navigation in cranio-maxillofacial surgery. *Oral and Maxillofacial Surgery Clinics of North America*, 22(1), 135–156.

Bose, S., Vahabzadeh, S., & Bandyopadhyay, A. (2013). Bone tissue engineering using 3D printing. *Materials Today*, 16(12), 496–504.

Choi, J. W., & Kim, N. (2015). Clinical application of three-dimensional printing technology in craniofacial plastic surgery. *Archives of Plastic Surgery*, 42(3), 267–277.

Ciocca, L., et al. (2010). CAD/CAM bilateral condylar prostheses for TMJ reconstruction. *International Journal of Oral and Maxillofacial Surgery*, 39(11), 1101–1108.

Ciocca, L., et al. (2012). CAD/CAM guided secondary mandibular reconstruction with free fibula flap and immediate implant rehabilitation. *Journal of Cranio-Maxillofacial Surgery*, 40(8), e511–e515.

D'haese, J., et al. (2017). Current state of the art of computer-guided implant surgery. *Periodontology* 2000, 73(1), 121–133.

Daly, A. C., et al. (2016). A comparison of different bioinks for 3D bioprinting of fibrocartilage and hyaline cartilage. *Biofabrication*, 8(4), 045002.

Dawood, A., Marti, B. M., Sauret-Jackson, V., & Darwood, A. (2015). 3D printing in dentistry. *British Dental Journal*, 219(11), 521–529.

Eggbeer, D., et al. (2012). Evaluation of direct and indirect additive manufacture of maxillofacial prostheses. *Proceedings of the Institution of Mechanical Engineers, Part H: Journal of Engineering in Medicine*, 226(9), 718–728.

Ellis, E., III, & Graham, J. (2002). Use of a 2.4-mm locking reconstruction plate/screw system for mandibular surgery. *Journal of Oral and Maxillofacial Surgery*, 60(6), 642–645.

Foley, B. D., et al. (2010). Mandibular reconstruction using computer-aided design and computer-aided manufacturing. *Oral and Maxillofacial Surgery Clinics of North America*, 22(1), 135–156.

George, E., et al. (2017). Measuring and establishing the accuracy and reproducibility of 3D printed medical models. *RadioGraphics*, 37(5), 1424–1450.

Giannopoulos, A. A., et al. (2016). Applications of 3D printing in cardiovascular diseases. *Nature Reviews Cardiology*, 13(12), 701–718.

Gibson, I., Rosen, D. W., & Stucker, B. (2015). *Additive manufacturing technologies*. Springer.

Grayson, W. L., et al. (2010). Engineering anatomically shaped human bone grafts. *Proceedings of the National Academy of Sciences of the United States of America*, 107(8), 3299–3304.

Gross, B. C., et al. (2014). Evaluation of 3D printing and its potential impact on biotechnology and the chemical sciences. *Analytical Chemistry*, 86(7), 3240–3253.

Gungor-Ozkerim, P. S., et al. (2018). Biopinks for 3D bioprinting. *Advanced Healthcare Materials*, 7(16), e1800700.

Hazeveld, A., Huddleston Slater, J. J., & Ren, Y. (2014). Accuracy and reproducibility of dental replica models reconstructed by different rapid prototyping techniques. *American Journal of Orthodontics and Dentofacial Orthopedics*, 145(1), 108–115.

Hull, C. W. (1986). Apparatus for production of three-dimensional objects by stereolithography (U.S. Patent No. 4,575,330).

Jardini, A. L., et al. (2014). Cranial reconstruction: 3D biomodel and custom-built implant created using additive manufacturing. *Journal of Cranio-Maxillofacial Surgery*, 42(8), 1877–1884.

Javaid, M., & Haleem, A. (2018). Additive manufacturing applications in medical cases: A literature-based review. *Alexandria Journal of Medicine*, 54(4), 411–422.

Javaid, M., & Haleem, A. (2018). Additive manufacturing applications in orthopaedics: A review. *Journal of Clinical Orthopaedics and Trauma*, 9(3), 202–206.

Javaid, M., & Haleem, A. (2019). Current status and applications of additive manufacturing in dentistry: A literature-based review. *Journal of Oral Biology and Craniofacial Research*, 9(3), 179–185.

Javaid, M., & Haleem, A. (2019). Industry 4.0 applications in medical field. *Clinical Epidemiology and Global Health*, 7(2), 317–321.

Kaigler, D., et al. (2013). Stem cell therapy for craniofacial bone regeneration. *Journal of Dental Research*, 92(7 Suppl), 113S–121S.

Kang, H. W., et al. (2016). A 3D bioprinting system to produce human-scale tissue constructs. *Nature Biotechnology*, 34(3), 312–319.

Kolesky, D. B., et al. (2014). 3D bioprinting of vascularized tissues. *Proceedings of the National Academy of Sciences of the United States of America*, 111(27), 12329–12334.

Kozakiewicz, M., et al. (2009). Clinical application of 3D pre-bent titanium implants for orbital floor reconstruction. *Journal of Cranio-Maxillofacial Surgery*, 37(4), 229–234.

Lim, K. H., et al. (2016). Use of 3D printed models in medical education. *Anatomical Sciences Education*, 9(6), 583–590.

Litjens, G., et al. (2017). A survey on deep learning in medical image analysis. *Medical Image Analysis*, 42, 60–88.

Mandrycky, C., et al. (2016). 3D bioprinting for engineering complex tissues. *Biotechnology Advances*, 34(4), 422–434.

Mangano, F., et al. (2017). Intraoral scanners in dentistry: A review of the current literature. *BMC Oral Health*, 17(1), 149.

Martelli, N., et al. (2016). Advantages and disadvantages of 3-dimensional printing in surgery: A systematic review. *Surgery*, 159(6), 1485–1500.

Martelli, N., et al. (2016). Advantages and disadvantages of 3-dimensional printing in surgery. *Surgery*, 159(6), 1485–1500.

Mazzoni, S., et al. (2013). Prosthetically guided maxillofacial reconstruction. *Journal of Oral and Maxillofacial Surgery*, 71(6), 1135–1145.

Mazzoni, S., et al. (2015). Computer-aided design and computer-aided manufacturing cutting guides and customized titanium plates are useful in upper maxilla waferless repositioning. *Journal of Oral and Maxillofacial Surgery*, 73(4), 701–707.

McMenamin, P. G., et al. (2014). The production of anatomical teaching resources using 3D printing technology. *Anatomical Sciences Education*, 7(6), 479–486.

Metzger, M. C., et al. (2007). Verification of clinical precision after computer-aided reconstruction in craniomaxillofacial surgery. *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology*, 104(1), e1–e10.

Mitsouras, D., et al. (2015). Medical 3D printing for the radiologist. *RadioGraphics*, 35(7), 1965–1988.

Modabber, A., et al. (2012). Evaluation of computer-assisted jaw reconstruction with free vascularized fibular flap compared to conventional surgery. *Journal of Cranio-Maxillofacial Surgery*, 40(8), 670–675.

Modabber, A., et al. (2012). Evaluation of computer-assisted jaw reconstruction with vascularized fibular flap. *Journal of Oral and Maxillofacial Surgery*, 70(5), e140–e147.

Momeni, F., et al. (2017). A review of 4D printing. *Materials & Design*, 122, 42–79.

Murphy, S. V., & Atala, A. (2014). 3D bioprinting of tissues and organs. *Nature Biotechnology*, 32(8), 773–785.

Murphy, S. V., & Atala, A. (2014). 3D bioprinting of tissues and organs. *Nature Biotechnology*, 32(8), 773–785.

Murphy, S. V., De Coppi, P., & Atala, A. (2020). Opportunities and challenges of translational 3D bioprinting. *Nature Biomedical Engineering*, 4(4), 370–380.

Murr, L. E., et al. (2010). Next-generation biomedical implants using additive manufacturing of complex, cellular and functional mesh arrays. *Philosophical Transactions of the Royal Society A: Mathematical, Physical and Engineering Sciences*, 368(1917), 1999–2032.

Oryan, A., et al. (2014). Bone regenerative medicine: Classic options, novel strategies, and future directions. *Journal of Orthopaedic Surgery and Research*, 9(1), 18.

Pittenger, M. F., et al. (1999). Multilineage potential of adult human mesenchymal stem cells. *Science*, 284(5411), 143–147.

Rasperini, G., et al. (2015). 3D-printed bioresorbable scaffold for periodontal repair. *Journal of Dental Research*, 94(9 Suppl), 153S–157S.

Rengier, F., et al. (2010). 3D printing based on imaging data. *International Journal of Computer Assisted Radiology and Surgery*, 5(4), 335–341.

Reymus, M., et al. (2019). 3D printed replicas for endodontic education. *International Endodontic Journal*, 52(1), 123–130.

Rose, A. S., et al. (2015). Multi-material 3D models for temporal bone surgical simulation. *Annals of Otology, Rhinology & Laryngology*, 124(7), 528–536.

Roser, S. M., et al. (2010). The accuracy of virtual surgical planning in free fibula mandibular reconstruction. *International Journal of Oral and Maxillofacial Surgery*, 39(3), 214–219.

Schneider, D., et al. (2009). A systematic review on accuracy and clinical outcome of computer-guided template-based implant dentistry. *Clinical Oral Implants Research*, 20(Suppl. 4), 73–86.

Schwendicke, F., Samek, W., & Krois, J. (2020). Artificial intelligence in dentistry: Chances and challenges. *Journal of Dental Research*, 99(7), 769–774.

Strbac, G. D., et al. (2017). Guided modern endodontic surgery: A novel approach for guided osteotomy and root resection. *Journal of Endodontics*, 43(3), 496–501.

Swennen, G. R., Mollemans, W., & Schutyser, F. (2009). Three-dimensional treatment planning of orthognathic surgery in the era of virtual imaging. *Journal of Oral and Maxillofacial Surgery*, 67(10), 2080–2092.

Tack, P., et al. (2016). 3D-printing techniques in a medical setting. *BioMedical Engineering OnLine*, 15(1), 115.

Tahayeri, A., et al. (2018). 3D printed versus conventionally cured provisional crown and bridge dental materials. *Dental Materials*, 34(2), 192–200.

Tahmaseb, A., et al. (2014). Computer technology applications in surgical implant dentistry. *International Journal of Oral and Maxillofacial Implants*, 29(Suppl), 25–42.

Toto, J. M., et al. (2015). Improved operative efficiency of free fibula flap mandible reconstruction with patient-specific, computer-guided preoperative planning. *Head & Neck*, 37(11), 1660–1664.

Van Assche, N., et al. (2012). Accuracy of computer-aided implant placement. *Clinical Oral Implants Research*, 23(Suppl. 6), 112–123.

Van Noort, R. (2012). The future of dental devices is digital. *Dental Materials*, 28(1), 3–12.

Ventola, C. L. (2014). Medical applications for 3D printing: Current and projected uses. *P & T*, 39(10), 704–711.

Vercruyssen, M., Laleman, I., Jacobs, R., & Quirynen, M. (2014). Computer-supported implant planning and guided surgery. *Periodontology 2000*, 66(1), 69–76.

Webster, T. J., & Ejiofor, J. U. (2004). Increased osteoblast adhesion on nanophase metals. *Biomaterials*, 25(19), 4731–4739.

Wilde, F., et al. (2015). Multicenter study on the use of patient-specific CAD/CAM reconstruction plates for mandibular reconstruction. *International Journal of Computerized Dentistry*, 18(3), 203–215.

Winder, J., & Bibb, R. (2005). Medical rapid prototyping technologies: State of the art and current limitations for application in oral and maxillofacial surgery. *Journal of Oral and Maxillofacial Surgery*, 63(7), 1006–1015.

Witowski, J., et al. (2018). From ideas to long-term studies: 3D printing clinical trials review. *International Journal of Computer Assisted Radiology and Surgery*, 13(9), 1473–1478.

Wolford, L. M., et al. (2015). Twenty-year follow-up study on a patient-fitted temporomandibular joint prosthesis. *Journal of Oral and Maxillofacial Surgery*, 73(5), 952–960.

Xia, J. J., Gateno, J., & Teichgraber, J. F. (2009). New clinical protocol to evaluate craniofacial deformity and plan surgical correction. *Journal of Oral and Maxillofacial Surgery*, 67(10), 2093–2106.

Yang, W. F., et al. (2018). Three-dimensional printing of patient-specific plates in oral and maxillofacial surgery. *Oral Oncology*, 78, 31–36.

Zhang, Y., et al. (2018). Finite element analysis and topology optimization of customized mandibular reconstruction plates. *Computer Methods in Biomechanics and Biomedical Engineering*, 21(15), 879–887.

Zinser, M. J., et al. (2014). Computer-assisted orthognathic surgery. *Oral and Maxillofacial Surgery Clinics of North America*, 26(4), 587–598.

Zweifel, D. F., et al. (2014). Computer-assisted reconstruction of maxillofacial defects using patient-specific implants. *International Journal of Oral and Maxillofacial Surgery*, 43(9), 1080–1087.

Zweifel, D. F., et al. (2015). Are virtual planning and guided surgery for head and neck reconstruction economically viable? *Journal of Oral and Maxillofacial Surgery*, 73(1), 170–175.

Zweifel, D. F., et al. (2015). Are virtual planning and patient-specific implants cost-effective? *Journal of Cranio-Maxillofacial Surgery*, 43(2), 170–175.

CHAPTER 3

THE ROLE OF VITAMIN D IN BONE METABOLISM: FROM MOLECULAR FOUNDATIONS TO THERAPEUTIC APPLICATIONS IN ORAL SURGERY

İsmail Faruk OKUMUŞ¹
Gökhan GÜRSES²

Introduction

The skeletal system is a metabolically active tissue that undergoes dynamic remodeling throughout life, requiring numerous hormonal and biochemical signals for this process to function properly.

In terms of chemical structure, vitamin D is not a single compound but a family of molecules comprising six distinct forms ranging from D2 to D7. However, since only vitamin D2 (ergocalciferol) and vitamin D3 (cholecalciferol) have demonstrated biological activity, the term "vitamin D" in the literature generally

¹ Research Assistant, Selçuk University Faculty of Dentistry, Department of Oral and Maxillofacial Surgery, ismail.okumus@selcuk.edu.tr, [0000-0002-2649-362X](tel:0000-0002-2649-362X)

² Doç Dr., Selçuk University Faculty of Dentistry, Department of Oral and Maxillofacial, gokhangurses.akademik@gmail.com, [0000-0002-3825-4650](tel:0000-0002-3825-4650)

refers to these two compounds (Kubodera, 2009). Vitamin D can be obtained through sun exposure, diet, and supplementation. Ultraviolet B radiation from sunlight at wavelengths of 290–315 nm penetrates the skin and converts 7-dehydrocholesterol first to previtamin D3 and then rapidly to vitamin D3. A diet rich in fatty fish is known to be effective in preventing vitamin D deficiency (Holick, 2007).

Both forms undergo a similar activation process once they enter the body. In the first step, 25-hydroxylation in the liver converts vitamin D to 25-hydroxyvitamin D [25(OH)D]. Unlike the second hydroxylation step in the kidney, this step is not subject to strict feedback regulation, allowing 25(OH)D to remain stable in the blood for extended periods by forming a complex with vitamin D-binding protein (DBP). Therefore, serum 25(OH)D level is considered the most reliable indicator of the body's vitamin D reserves (Kubodera, 2009). When a physiological need arises, 25(OH)D is taken up by proximal renal tubular cells via the megalin protein and converted by the 1 α -hydroxylase enzyme to 1,25-dihydroxyvitamin D (calcitriol), the most biologically potent form (Kubodera, 2009). Calcitriol binds to vitamin D receptors (VDR) in target organs such as the small intestine, bone tissue, kidneys, and parathyroid glands to initiate various biological responses (Kubodera, 2009).

Vitamin D stimulates osteoclastogenesis by directing osteoblasts to produce RANKL (receptor activator of nuclear factor- κ B ligand) and activates resting osteoclasts for bone resorption (Suda et al., 2003).

Vitamin D deficiency was once considered a largely resolved issue following food fortification with vitamin D and the control of rickets; however, this assumption has since proven inadequate. Rickets is merely the most prominent manifestation of vitamin D deficiency, and the problem extends far beyond this disease. Indeed, vitamin D deficiency predisposes to growth retardation, skeletal

deformities, and increased risk of hip fracture in later life during childhood, and to osteopenia, osteoporosis, osteomalacia, and muscle weakness in adults (Holick, 2007). Research has revealed that the effects of 1,25-dihydroxyvitamin D₃ are not limited to calcium and phosphate homeostasis. This compound has been found to regulate numerous cellular processes, including cell growth and differentiation, innate and adaptive immune function, cardiovascular function, and complex interactions with other hormones; its potential role in cancer, cardiovascular disease, and autoimmune conditions has attracted intense research interest (Christakos et al., 2016).

This chapter aims to provide a comprehensive perspective on the role of vitamin D in bone metabolism, addressing molecular mechanisms, clinical evidence, and therapeutic applications.

Molecular Mechanisms

Following 1 α ,25(OH)₂D₃ binding, the nuclear VDR heterodimerizes with Retinoid X Receptor (RXR). The resulting dimeric VDR complex binds to genomic DNA at the regulatory regions of primary vitamin D target genes (Carlberg & Molnár, 2015; Haussler et al., 2013). VDR expression is regulated by 1 α ,25(OH)₂D₃ itself—via high-affinity and high-specificity binding to the ligand-binding pocket (Maestro et al., 2016)—and by various other factors including PTH, glucocorticoids, TGF β , and EGF (Godschalk et al., 1992; Reinhardt & Horst, 1990; van Leeuwen et al., 1992). The VDR promoter region contains multifunctional enhancers that regulate gene transcription (Zella et al., 2010).

The role of the vitamin D receptor (VDR) in bone metabolism has been largely elucidated through gene deletion mouse models. In mice lacking a functional 1,25(OH)₂D/VDR system, the number of osteoblasts was found to be increased, but the increase in bone volume was largely due to accumulation of unmineralized osteoid (Goltzman, 2018; Li et al., 1997). Treatment of these mice

with 1,25(OH)₂D₃ normalized mineral levels and PTH, and fully restored bone architecture (Dardenne et al., 2003; Goltzman, 2018). These findings indicate that the 1,25(OH)₂D/VDR system is indispensable for normal bone formation.

The effect of VDR on bone is not limited to mineralization. In transgenic mouse models in which VDR expression was examined in late osteoblasts and early osteocytes, a marked increase in cortical and trabecular bone, increased periosteal bone formation, and decreased bone resorption were observed (Gardiner et al., 2000; Triliana et al., 2016). This anti-resorptive effect is thought to occur through a reduction in the RANKL/OPG ratio, while the anabolic effect is believed to be mediated by increased expression of LRP-5, which functions as a co-receptor in the Wnt signaling pathway (Fretz et al., 2007). Therefore, the 1,25(OH)₂D/VDR system may exert anabolic effects on bone tissue through its actions on late osteoblastic and osteocytic cells (Goltzman, 2018).

The molecular interaction between RUNX2, a master transcription factor governing osteoblast development, and VDR is considered central to mediating the effects of vitamin D on osteoblast differentiation and mineralization. Evidence suggests that this interaction modulates the gene expression of two non-collagenous matrix proteins, osteocalcin and osteopontin, in rat osteoblasts (Paredes et al., 2004; Shen & Christakos, 2005). Furthermore, VDR that has been activated by 1 α ,25(OH)₂D₃ is also known to engage with the Wnt signaling pathway, thereby contributing to the regulation of osteoblast differentiation (Haussler et al., 2010). The interplay between vitamin D and the gene networks governing osteoblast differentiation and bone formation has been thoroughly examined and discussed in a comprehensive review by van de Peppel and van Leeuwen (van de Peppel & van Leeuwen, 2014).

Beyond its direct actions on bone tissue, vitamin D is also recognized for its indirect role in supporting skeletal integrity through the regulation of calcium homeostasis. Among its principal postnatal functions is the maintenance of calcium balance, achieved primarily by enhancing intestinal calcium absorption, especially under conditions of hypocalcemia. In states of low dietary calcium intake, 1,25(OH)₂D promotes transcellular calcium transport predominantly in the duodenum, while also acting on more distal portions of the intestinal tract. This response is predominantly mediated through the upregulation of the apical membrane calcium channel TRPV6 and the calcium-binding protein calbindin-D9k. Conversely, when calcium intake is abundant, paracellular calcium transport gains greater significance, and this alternative route remains subject to regulation by 1,25(OH)₂D (Goltzman, 2018).

The effects of 1,25(OH)₂D are not limited to slow genomic responses involving gene transcription; this molecule also has effects that develop too rapidly to require a genomic mechanism. These non-genomic effects were first described in the context of rapid stimulation of intestinal calcium transport—transcaltachia—in vitamin D-replete chicks, and were subsequently extended to effects on growth plate chondrocytes and skin keratinocytes (Bikle, 2014). The molecular basis of these rapid responses focuses on two receptors: VDR itself and the membrane-associated rapid response steroid-binding protein MARRS (also known as ERp57/GRp58/ERp60). Both receptors have been reported to be localized in membrane caveolae/lipid rafts and to be poised to activate kinases, phosphatases, and ion channels (Bikle, 2014). It has been proposed that genomic VDR agonists bind in the 6-s-trans configuration, whereas agonists specific to rapid responses are in the 6-s-cis configuration and trigger these rapid responses through a VDR model with an alternative ligand pocket (Bikle, 2014).

Clinical Evidence

The molecular effects of vitamin D on bone metabolism have important clinical implications in the oral and maxillofacial region as well. Evidence is increasingly supporting a direct association between vitamin D levels and key clinical outcomes such as alveolar bone homeostasis, dental implant osseointegration success, and periodontal tissue integrity.

Periodontitis is characterized as a multifactorial infectious condition that leads to the progressive destruction of periodontal tissues through a complex pathogenic process (Chen et al., 2023). Clinical data indicate that vitamin D deficiency is directly associated with the risk of periodontitis (Anbarcioglu et al., 2019; Isola et al., 2020). The effects of 1,25 D₃ on alveolar bone are mediated through multiple mechanisms, including immunomodulatory effects, inhibition of osteoclastogenesis, induction of osteogenic differentiation, and transcriptional regulation of osteogenesis-related factors (Chen et al., 2023). These mechanisms have been reflected in clinical data. A cross-sectional study conducted on 562 elderly men reported that daily vitamin D intake below 400 IU was associated with increased alveolar bone resorption, whereas daily intake above 800 IU significantly reduced the risk of severe chronic periodontitis (Alshouibi et al., 2013). These findings suggest that adequate vitamin D levels play a critical role in maintaining alveolar bone homeostasis.

These effects of vitamin D on alveolar bone are also of great significance for dental implant osseointegration. A systematic review examining the effect of vitamin D deficiency on dental implant osseointegration reported that findings from all animal studies showed decreased bone-implant contact (BIC) and new bone formation around implants. Furthermore, the majority of studies reviewed showed that systemic vitamin D supplementation increased new bone formation around implants (Werny et al., 2022). The beneficial effect of vitamin D supplementation on

osseointegration was more pronounced in individuals with systemic diseases such as diabetes, osteoporosis, and chronic kidney disease (Werny et al., 2022). Data derived from clinical studies suggest that individuals presenting with inadequate serum vitamin D levels may face an elevated risk of early implant failure; however, the existing literature on the influence of vitamin D supplementation on osseointegration in clinical practice is still insufficient, underscoring the necessity for more comprehensive and well-designed studies (Werny et al., 2022).

When the existing body of evidence is considered collectively, it becomes apparent that vitamin D exerts both direct and indirect influences on bone metabolism within the oral and maxillofacial region. Vitamin D status appears to serve a determinative function across a broad clinical spectrum, encompassing conditions that range from periodontal bone loss to implant osseointegration. Nevertheless, given that the majority of existing studies are based on animal models and that the level of evidence in human studies remains limited, comprehensive, randomized controlled clinical trials aimed at determining optimal vitamin D levels and supplementation protocols are needed.

Therapeutic Potential and Clinical Applications

Evidence from both animal models and human studies suggests that maintaining adequate vitamin D levels may enhance osseointegration by positively influencing the quality and quantity of newly formed bone at the implant-tissue interface, ultimately contributing to more predictable and favorable clinical outcomes (Sundar et al., 2023). Vitamin D supplementation has also been shown to support bone defect regeneration, markedly increasing new bone formation and mineralization, particularly in vitamin D-deficient patients (Sundar et al., 2023).

Although supplementation protocols vary across studies and no single approach has yet been universally validated, current findings indicate that careful management of vitamin D levels may be a critical factor in improving osseointegration and implant success. Animal studies, which are more prevalent than human research, demonstrate that systemic vitamin D supplementation increases new bone formation around implants, though this finding is not universal (Wu et al., 2013; Zhou et al., 2012).

Achieving adequate serum 25(OH)D levels preoperatively is of great importance for successful osseointegration in dental implant surgery. In a prospective, randomized controlled clinical trial conducted by Kwiatek et al. (2021), 25(OH)D reference values were classified as deficient (<20 ng/mL), low (20–30 ng/mL), optimal (30–50 ng/mL), high (50–100 ng/mL), and toxic (>150 ng/mL). The study findings revealed a positive association between 25(OH)D level on the day of surgery and the bone level at the implant site; patients with serum levels ≥ 30 ng/mL had significantly higher bone levels at the implant site at postoperative weeks 6 and 12. Additionally, in the patient group with preoperative 25(OH)D levels <30 ng/mL who received daily supplementation of 8,000 IU vitamin D in accordance with European guidelines, bone levels at week 12 were statistically significantly higher compared to the group that received no supplementation (Kwiatek et al., 2021). These findings suggest that evaluating serum 25(OH)D levels prior to implant surgery and initiating supplementation therapy when necessary may be of clinical importance.

The effects of vitamin D on bone metabolism cannot be considered independently of its synergistic relationship with calcium. Vitamin D increases calcium absorption in the small intestine, thereby maintaining adequate serum calcium levels and indirectly contributing to bone mineralization (Christakos et al., 2016; Holick, 2007). Although it has been observed in animal

models that calcium supplementation combined with vitamin D supplementation may provide additional benefit to alveolar bone levels, human studies in this area remain insufficient (Hong et al., 2015).

Conclusion

Vitamin D emerges as a prohormone that plays a central role in the regulation of bone metabolism, exerting multifaceted effects on osteoblast differentiation, osteoclastogenesis, and calcium homeostasis through genomic and non-genomic pathways. At the molecular level, its effects on VDR-mediated signaling and calcium-phosphate metabolism are manifested at the macroscopic level as bone mineral density and structural integrity. In the context of the oral and maxillofacial region specifically, vitamin D levels appear to play a determinant role in alveolar bone homeostasis, periodontal tissue health, and dental implant osseointegration.

Nevertheless, it should not be overlooked that a significant portion of the existing literature is based on animal models and that the level of evidence in human studies remains insufficient. Determining the optimal preoperative serum 25(OH)D level for implant surgery, establishing standardized supplementation protocols, and investigating the long-term effects of combined calcium and vitamin D therapy in the oral context should constitute the primary agenda items for future research. High-quality, large-sample, long-term follow-up randomized controlled clinical trials are needed in this field.

In conclusion, incorporating assessment of vitamin D status into routine clinical evaluation and planning supplementation therapy when indicated stands out as an important approach for clinicians seeking to support implant success and overall bone health.

REFERENCES

Alshouibi, E. N., Kaye, E. K., Cabral, H. J., Leone, C. W. & Garcia, R. I. (2013). Vitamin D and periodontal health in older men. *Journal of Dental Research*, 92(8), 689–693. <https://doi.org/10.1177/0022034513495239>

Anbarcioglu, E., Kirtiloglu, T., Öztürk, A., Kolbakir, F., Acıkgöz, G. & Colak, R. (2019). Vitamin D deficiency in patients with aggressive periodontitis. *Oral Diseases*, 25(1), 242–249. <https://doi.org/10.1111/odi.12968>

Bikle, D. D. (2014). Vitamin D metabolism, mechanism of action, and clinical applications. *Chemistry & Biology*, 21(3), 319–329. <https://doi.org/10.1016/j.chembiol.2013.12.016>

Carlberg, C. & Molnár, F. (2015). Vitamin D receptor signaling and its therapeutic implications: Genome-wide and structural view. *Canadian Journal of Physiology and Pharmacology*, 93(5), 311–318. <https://doi.org/10.1139/cjpp-2014-0383>

Chen, X., Arias, Z., Omori, K., Yamamoto, T., Shinoda-Ito, Y. & Takashiba, S. (2023). Autophagy as a potential mechanism underlying the biological effect of 1,25-Dihydroxyvitamin D3 on periodontitis: a narrative review. *BMC Oral Health*, 23(1), 90. <https://doi.org/10.1186/s12903-023-02802-9>

Christakos, S., Dhawan, P., Verstuyf, A., Verlinden, L. & Carmeliet, G. (2016). Vitamin D: Metabolism, molecular mechanism of action, and pleiotropic effects. *Physiological Reviews*, 96(1), 365–408. <https://doi.org/10.1152/physrev.00014.2015>

Dardenne, O., Prudhomme, J., Hacking, S. A., Glorieux, F. H. & St-Arnaud, R. (2003). Rescue of the pseudo-vitamin D deficiency rickets phenotype of CYP27B1-deficient mice by treatment with 1,25-dihydroxyvitamin D3: biochemical, histomorphometric, and biomechanical analyses. *Journal of Bone*

and Mineral Research: The Official Journal of the American Society for Bone and Mineral Research, 18(4), 637–643.
<https://doi.org/10.1359/jbmr.2003.18.4.637>

Fretz, J. A., Zella, L. A., Kim, S., Shevde, N. K. & Pike, J. W. (2007). 1,25-Dihydroxyvitamin D₃ induces expression of the Wnt signaling co-regulator LRP5 via regulatory elements located significantly downstream of the gene's transcriptional start site. *The Journal of Steroid Biochemistry and Molecular Biology*, 103(3-5), 440–445. <https://doi.org/10.1016/j.jsbmb.2006.11.018>

Gardiner, E. M., Baldock, P. A., Thomas, G. P., Sims, N. A., Henderson, N. K., Hollis, B., White, C. P., Sunn, K. L., Morrison, N. A., Walsh, W. R. & Eisman, J. A. (2000). Increased formation and decreased resorption of bone in mice with elevated vitamin D receptor in mature cells of the osteoblastic lineage. *FASEB Journal: Official Publication of the Federation of American Societies for Experimental Biology*, 14(13), 1908–1916.
<https://doi.org/10.1096/fj.99-1075com>

Godschalk, M., Levy, J. R. & Downs, R. W., Jr. (1992). Glucocorticoids decrease vitamin D receptor number and gene expression in human osteosarcoma cells. *Journal of Bone and Mineral Research: The Official Journal of the American Society for Bone and Mineral Research*, 7(1), 21–27.
<https://doi.org/10.1002/jbmr.5650070105>

Goltzman, D. (2018). Functions of vitamin D in bone. *Histochemistry and Cell Biology*, 149(4), 305–312.
<https://doi.org/10.1007/s00418-018-1648-y>

Haussler, M. R., Haussler, C. A., Whitfield, G. K., Hsieh, J.-C., Thompson, P. D., Barthel, T. K., Bartik, L., Egan, J. B., Wu, Y., Kubicek, J. L., Lowmiller, C. L., Moffet, E. W., Forster, R. E. & Jurutka, P. W. (2010). The nuclear vitamin D receptor controls the expression of genes encoding factors which feed the “Fountain of

Youth” to mediate healthful aging. *The Journal of Steroid Biochemistry and Molecular Biology*, 121(1-2), 88–97. <https://doi.org/10.1016/j.jsbmb.2010.03.019>

Haussler, M. R., Whitfield, G. K., Kaneko, I., Haussler, C. A., Hsieh, D., Hsieh, J.-C. & Jurutka, P. W. (2013). Molecular mechanisms of vitamin D action. *Calcified Tissue International*, 92(2), 77–98. <https://doi.org/10.1007/s00223-012-9619-0>

Holick, M. F. (2007). Vitamin D deficiency. *The New England Journal of Medicine*, 357(3), 266–281. <https://doi.org/10.1056/NEJMra070553>

Hong, H.-H., Yen, T.-H., Hong, A. & Chou, T.-A. (2015). Association of vitamin D3 with alveolar bone regeneration in dogs. *Journal of Cellular and Molecular Medicine*, 19(6), 1208–1217. <https://doi.org/10.1111/jcmm.12460>

Isola, G., Alibrandi, A., Rapisarda, E., Matarese, G., Williams, R. C. & Leonardi, R. (2020). Association of vitamin D in patients with periodontitis: A cross-sectional study. *Journal of Periodontal Research*, 55(5), 602–612. <https://doi.org/10.1111/jre.12746>

Kubodera, N. (2009). A new look at the most successful prodrugs for active vitamin D (D hormone): alfacalcidol and doxercalciferol. *Molecules (Basel, Switzerland)*, 14(10), 3869–3880. <https://doi.org/10.3390/molecules14103869>

Kwiatek, J., Jaroń, A. & Trybek, G. (2021). Impact of the 25-hydroxycholecalciferol concentration and vitamin D deficiency treatment on changes in the bone level at the implant site during the process of osseointegration: A prospective, randomized, controlled clinical trial. *Journal of Clinical Medicine*, 10(3), 526. <https://doi.org/10.3390/jcm10030526>

Li, Y. C., Pirro, A. E., Amling, M., Delling, G., Baron, R., Bronson, R. & Demay, M. B. (1997). Targeted ablation of the vitamin D receptor: an animal model of vitamin D-dependent rickets type II with alopecia. *Proceedings of the National Academy of Sciences of the United States of America*, 94(18), 9831–9835. <https://doi.org/10.1073/pnas.94.18.9831>

Maestro, M. A., Molnár, F., Mouriño, A. & Carlberg, C. (2016). Vitamin D receptor 2016: novel ligands and structural insights. *Expert Opinion on Therapeutic Patents*, 26(11), 1291–1306. <https://doi.org/10.1080/13543776.2016.1216547>

Paredes, R., Arriagada, G., Cruzat, F., Villagra, A., Olate, J., Zaidi, K., van Wijnen, A., Lian, J. B., Stein, G. S., Stein, J. L. & Montecino, M. (2004). Bone-specific transcription factor Runx2 interacts with the 1 α ,25-dihydroxyvitamin D₃ receptor to up-regulate rat osteocalcin gene expression in osteoblastic cells. *Molecular and Cellular Biology*, 24(20), 8847–8861. <https://doi.org/10.1128/MCB.24.20.8847-8861.2004>

Reinhardt, T. A. & Horst, R. L. (1990). Parathyroid hormone down-regulates 1,25-dihydroxyvitamin D receptors (VDR) and VDR messenger ribonucleic acid in vitro and blocks homologous up-regulation of VDR in vivo. *Endocrinology*, 127(2), 942–948. <https://doi.org/10.1210/endo-127-2-942>

Shen, Q. & Christakos, S. (2005). The vitamin D receptor, Runx2, and the Notch signaling pathway cooperate in the transcriptional regulation of osteopontin. *The Journal of Biological Chemistry*, 280(49), 40589–40598. <https://doi.org/10.1074/jbc.M504166200>

Suda, T., Ueno, Y., Fujii, K. & Shinki, T. (2003). Vitamin D and bone: Vitamin D and Bone. *Journal of Cellular Biochemistry*, 88(2), 259–266. <https://doi.org/10.1002/jcb.10331>

Sundar, R., Bhagavandas Rai, A., Naveen Kumar, J. & Devang Divakar, D. (2023). The role of Vitamin D as an adjunct for bone regeneration: A systematic review of literature. *The Saudi Dental Journal*, 35(3), 220–232. <https://doi.org/10.1016/j.sdentj.2023.02.002>

Triliana, R., Lam, N. N., Sawyer, R. K., Atkins, G. J., Morris, H. A. & Anderson, P. H. (2016). Skeletal characterization of an osteoblast-specific vitamin D receptor transgenic (ObVDR-B6) mouse model. *The Journal of Steroid Biochemistry and Molecular Biology*, 164, 331–336. <https://doi.org/10.1016/j.jsbmb.2015.08.009>

van de Peppel, J. & van Leeuwen, J. P. T. M. (2014). Vitamin D and gene networks in human osteoblasts. *Frontiers in Physiology*, 5, 137. <https://doi.org/10.3389/fphys.2014.00137>

van Leeuwen, J. P., Birkenhäger, J. C., Buurman, C. J., van den Bemd, G. J., Bos, M. P. & Pols, H. A. (1992). Bidirectional regulation of the 1,25-dihydroxyvitamin D₃ receptor by phorbol ester-activated protein kinase-C in osteoblast-like cells: interaction with adenosine 3',5'-monophosphate-induced up-regulation of the 1,25-dihydroxyvitamin D₃ receptor. *Endocrinology*, 130(4), 2259–2266. <https://doi.org/10.1210/endo.130.4.1312452>

Werny, J. G., Sagheb, K., Diaz, L., Kämmerer, P. W., Al-Nawas, B. & Schiegnitz, E. (2022). Does vitamin D have an effect on osseointegration of dental implants? A systematic review. *International Journal of Implant Dentistry*, 8(1), 16. <https://doi.org/10.1186/s40729-022-00414-6>

Wu, Y.-Y., Yu, T., Yang, X.-Y., Li, F., Ma, L., Yang, Y., Liu, X.-G., Wang, Y.-Y. & Gong, P. (2013). Vitamin D₃ and insulin combined treatment promotes titanium implant osseointegration in diabetes mellitus rats. *Bone*, 52(1), 1–8. <https://doi.org/10.1016/j.bone.2012.09.005>

Zella, L. A., Meyer, M. B., Nerenz, R. D., Lee, S. M., Martowicz, M. L. & Pike, J. W. (2010). Multifunctional enhancers regulate mouse and human vitamin D receptor gene transcription. *Molecular Endocrinology (Baltimore, Md.)*, 24(1), 128–147. <https://doi.org/10.1210/me.2009-0140>

Zhou, C., Li, Y., Wang, X., Shui, X. & Hu, J. (2012). 1,25Dihydroxy vitamin D(3) improves titanium implant osseointegration in osteoporotic rats. *Oral Surgery, Oral Medicine, Oral Pathology and Oral Radiology*, 114(5 Suppl), S174–S178. <https://doi.org/10.1016/j.oooo.2011.09.030>

CHAPTER 4

GRAFT MATERIALS AND MECHANISMS OF BONE FORMATION

1. Gökhan GÜRSES¹

2. İsmail Faruk OKUMUŞ²

Introduction

Bone defects of the craniomaxillofacial skeleton range from small (a few millimeters) periodontal defects to large segmental defects resulting from trauma, surgical excision, or cranioplasty. Such defects generally have complex three-dimensional structural requirements and are difficult to repair (Elsalanty & Genecov, 2009). In this context, bone grafting is the most widely accepted surgical approach for the repair of these defects. The need for bone graft materials in dentistry has increased significantly in recent years, driven by advances in dental implantology and the growing clinical demand for the repair of craniofacial bone defects (Zhao et al.,

¹ Doç Dr., Selçuk University Faculty of Dentistry, Department of Oral and Maxillofacial, gokhangurses.akademik@gmail.com, [0000-0002-3825-4650](tel:0000-0002-3825-4650)

²Research Assistant, Selçuk University Faculty of Dentistry, Department of Oral and Maxillofacial Surgery, ismail.okumus@selcuk.edu.tr, [0000-0002-2649-362X](tel:0000-0002-2649-362X)

2021) . A bone graft is defined as living tissue transplanted into a bone defect alone or in combination with other materials to support bone healing, promoting bone regeneration through the mechanisms of osteoconduction, osteoinduction, and osteogenesis (Ciszyński et al., 2023; Zhao et al., 2021). Bone grafts are classified into four main categories according to their source and biological properties: autograft, allograft, xenograft, and alloplastic (synthetic) materials (Miron, 2024). Among the current options in the field of bone regeneration, autogenous bone is the gold standard due to its osteoinductive and osteogenic capabilities. All other materials (allografts, xenografts, and synthetic biomaterials) have limitations that must be considered depending on their use (Ahmad Oryan et al., 2014).

Three fundamental ossification mechanisms — osteogenesis, osteoinduction, and osteoconduction — play a decisive role in the clinical success of graft materials, and understanding these mechanisms is of great importance for clinicians. The existing literature reports that the osteogenic potential of different graft types varies considerably depending on the preparation method, storage conditions, and patient factors (Ciszyński et al., 2023). In this context, a comprehensive analysis of the ossification mechanisms of graft materials will make important contributions to evidence-based decision-making in material selection by clinicians.

This review first addresses the mechanisms of bone formation — osteogenesis, osteoinduction, and osteoconduction — in terms of their biological foundations and cellular processes, then comparatively examines the ossification potential of the main graft material categories — autograft, allograft, xenograft, and alloplastic materials. Subsequently, biological factors affecting bone formation and the role of growth factors are discussed, and finally clinical application areas in dentistry are evaluated. This comprehensive approach aims to provide clinicians with an up-to-date and evidence-

based guide for graft material selection and optimization of the ossification process.

Mechanisms of Bone Formation

The regenerative potential of bone graft materials is built upon three fundamental biological mechanisms: osteogenesis, osteoinduction, and osteoconduction. Each of these mechanisms functions at different levels depending on the graft type and host conditions, and supports new bone formation in a complementary manner(Ciszyński et al., 2023; Zhao et al., 2021).

Osteogenesis

Osteogenesis is the process of new bone synthesis carried out by donor cells obtained from the host or graft donor. In this process, mesenchymal stem cells (MSCs), osteoblasts, and osteocytes play key roles(Khan et al., 2005). Typically, only fresh autologous grafts and bone marrow transplants — whether auto- or allografted — are used in this process(Roberts & Rosenbaum, 2012). Osteogenic capacity is considered the primary determinant of the superiority of autografts over other graft types, owing to the ability of viable osteoblasts within the graft to directly produce bone matrix at the implantation site(Haugen et al., 2026).

Osteoinduction

Osteoinduction refers to the conversion and differentiation of pluripotent stem cells into bone-forming cells through biochemical signaling pathways mediated by growth factors such as bone morphogenetic proteins (BMPs) (Thieu et al., 2021). Among the available graft materials, it has been reported that only autografts and allografts are capable of cell recruitment through chemotactic growth factors such as BMP-2 and PDGF, while xenografts and synthetic alloplasts lack this capacity (Miron, 2024). Growth factors including BMP-2, -4 and -7, as well as PDGF, FGF, interleukins,

and VEGF play key regulatory roles in the osteoinduction process (Roberts & Rosenbaum, 2012).

Osteoconduction

Osteoconduction provides the structural framework that guides bone growth and vascularization, requiring materials with appropriate porosity, surface chemistry, and mechanical properties (Thieu et al., 2021). In this process, the implanted scaffold passively allows the ingrowth of host capillaries, perivascular tissue, and mesenchymal stem cells, thereby laying the groundwork for new bone formation (Roberts & Rosenbaum, 2012). Although all graft materials by definition possess osteoconductive properties, this mechanism alone is not sufficient to initiate new bone formation (Miron, 2024). Allografts and xenografts exhibit osteoinductive and osteoconductive properties, but lack the osteogenic potential possessed by autografts (Ahmad Oryan et al., 2014).

Graft Materials and Their Osteogenic Potential

Bone graft materials are classified into four main categories according to their biological origin: autograft, allograft, xenograft, and alloplastic materials. Each category has its own unique advantages and limitations in terms of osteogenic potential, ease of clinical use, and biosafety profile (Ahmad Oryan et al., 2014).

Autograft

Bone grafts harvested from one site of the same individual and implanted into another site are referred to as autograft, autologous, or autogenous bone graft (Zimmermann & Moghaddam, 2011). These grafts can be obtained from both intraoral and extraoral donor sites such as the iliac crest, cranium, mandible, radius, and tibia, and can be applied in cancellous, cortical (avascular or vascularized), or corticocancellous form (Kämmerer & Al-

Nawas, 2023; Stevens et al., 2021). Intraoral donor sites offer significant advantages over extraoral sites in terms of ease of surgical access, proximity to donor and recipient sites, and less postoperative pain (Altıparmak et al., 2015). Fresh autografts contain viable cells along with osteoinductive proteins such as BMP-2, BMP-7, FGF, IGF, and PDGF (Nandi et al., 2010; Pereira-Júnior et al., 2007). Due to the presence of mesenchymal stem cells (MSCs), osteoprogenitor cells, osteogenic cells, and growth factors together, fresh autografts possess the most optimal level among all graft types in terms of osteogenic, osteoinductive, and osteoconductive properties (Keating & McQueen, 2001; A. Oryan et al., 2012). Autografts carry no risk of viral transmission; they also provide mechanical support to implanted structures and become incorporated into the surrounding bone through creeping substitution — a remodeling process involving gradual resorption with simultaneous new bone formation — eventually transforming into a mechanically efficient structure (Greenwald et al., 2001). However, autografts have important disadvantages including the need for a second surgical site, donor site morbidity, limited graft quantity, and increased operative time (Ferraz, 2023).

The mechanism of bone healing and osteogenesis in autografts varies depending on the graft type (Marx, 2007)[20]. In avascular block bone grafts, osteocytes die after transplantation due to their entrapment within the mineral matrix and disruption of canalicular blood circulation; new bone formation is largely achieved through osteoinduction and osteoconduction from adjacent bone edges (Marx, 2007). In vascularized grafts, mature osteocytes, periosteum, and the mineral matrix are transferred alive, so healing follows the same mechanism as fracture healing; the process initiated by platelet degranulation triggers the proliferation of endosteal and periosteal osteoblasts, leading to the formation of bone callus (Marx, 2007). Autogenous cancellous bone marrow

grafts are the graft type with the highest osteogenic potential; the transplanted osteocompetent cells survive through plasmatic circulation until revascularization is complete. This process begins with platelets releasing growth factors such as PDGF, TGF- β , VEGF, and EGF, and is maintained by macrophages until revascularization is completed between days 14 and 21 (Marx, 2007; Roberts & Rosenbaum, 2012).

Allograft

Allografts are bone grafts harvested from one individual and transplanted to another individual of the same species (Ehrler & Vaccaro, 2000). Considering the limitations encountered in obtaining autografts, allografts are widely used as an alternative to autografts in both clinical and experimental settings (Ehrler & Vaccaro, 2000). Allografts are graft materials that can be cortical, cancellous, or corticocancellous in structure and are made available in clinical use in various shapes and sizes. Fresh frozen bone (FFB), freeze-dried bone allograft (FDBA), and demineralized freeze-dried bone allograft (DFDBA) are among the most common forms of this graft type. These materials predominantly exhibit osteoconductive properties, but can also acquire osteoinductive potential depending on certain preparation methods (Sheikh et al., 2017). In allografts, the demineralization process enhances access to a large number of growth factors, primarily BMP-2, making DFDBAs superior in osteoinductive potential compared to FDBAs (Miron, 2024). The biology of healing and integration of allograft bone is similar to that of autografts, but the key difference is the absence of donor cells; therefore, the integration of both cancellous and cortical allografts proceeds more slowly than equivalent autografts (Goldberg & Stevenson, 1987). However, allografts have important disadvantages including the risk of immune response, potential disease transmission risk, and variable osteoinductivity depending on the

donor and preparation method (Miron, 2024; Ahmad Oryan et al., 2014).

Xenograft

Xenografts are graft materials produced from the inorganic components of bone obtained from different animal species, and their fundamental biological properties are characterized by osteoconduction and limited resorption capacity. In clinical practice, these materials are used in combination with growth factors or bone grafts of different origins to enhance their biological potential. Xenografts share several advantages with allografts, but their clinical efficacy remains a matter of debate. The existing literature demonstrates that, compared to allografts, xenografts show more pronounced connective tissue ingrowth, delayed revascularization, and a lower rate of resorption (Serrano Méndez et al., 2017). Commonly used xenografts are obtained from various sources including coral, porcine, and bovine origins (Develioglu et al., 2009; Ahmad Oryan et al., 2012). When processed to be made safe for human transplantation, xenogeneic bone grafts theoretically offer a source of unlimited potential (Develioglu et al., 2009).

Xenografts are prepared by various methods such as heat treatment, hydrothermal hydrolysis with NaOH, ethylenediamine, or sodium hypochlorite; the primary objective in these processes is to remove organic components, xenogeneic antigens, and cellular residues while preserving the material's natural biological properties (Jung et al., 2020; Long et al., 2012). The clinical efficacy of xenograft scaffolds depends on having ideal porosity, appropriate pore size, and a well-interconnected pore network. In addition, particle size and the calcium/phosphorus (Ca/P) ratio are of great importance for bone regeneration. The presence of these properties together creates an effective bone healing environment by supporting cell adhesion, proliferation and migration, the release of

bioactive molecules, and vascularization (Amid et al., 2021; El-Rashidy et al., 2017; Fan et al., 2018). However, xenografts have important disadvantages including the risk of immune response, delayed bone remodeling due to low resorption rates, and the risk of zoonotic disease transmission (Ahmad Oryan et al., 2014).

Alloplastic Materials

Interest in the development of osteoconductive matrices derived from synthetic porous structures as an alternative to biologically-derived materials for bone defect repair has increased significantly. A clinically effective synthetic material is expected to be able to mimic the architecture of host cancellous bone and provide a microenvironment suitable for the adhesion, integration, and proliferation of mesenchymal stem cells (MSCs). In this context, calcium phosphate (CaP) and calcium sulfate (CaS)-based compounds are among the most widely used synthetic bone graft substitutes in clinical practice (Baldwin et al., 2019).

Among the developed alloplastic materials are hydroxyapatite, coral- and algae-derived hydroxyapatite derivatives, calcium phosphate compounds, calcium sulfate, collagen, and polymers. With the exception of the P-15 peptide (a 15-amino acid synthetic peptide that mimics the cell-binding domain of type I collagen and is claimed to stimulate the differentiation of mesenchymal cells into osteoblasts), the vast majority of these synthetic materials are accepted as inert compounds devoid of or with extremely limited osteoinductive capacity (Kübler et al., 2004; Turhani et al., 2003, 2005). These materials, which can be prepared in granule, block, or paste format, are manufactured with high porosity to facilitate vascular penetration and distribution (Chappard, 2017; Lu et al., 2021).

The most widely used bioceramic material in human bone grafting is hydroxyapatite (HA), which closely resembles bone

mineral in terms of chemical composition and crystal structure (Supová, 2009). HA and some other calcium-based ceramic materials are accepted as bioactive materials because they have been reported to support bone growth (Heilmann et al., 2007; Martinetti et al., 2005). This bioactivity is directly related to its osteoconductive properties that allow osteoblasts to adhere to and migrate along the material surface (Frayssinet et al., 1998; Hench, 1991). Due to the slow and limited resorption potential of HA, the implanted material cannot be completely replaced with new bone; however, it can function as a volumetric filler (Bohner et al., 2020; Sheikh et al., 2017). In contrast, β -TCP is easily resorbed due to its interconnected porous structure and is rapidly replaced by newly formed bone tissue (Bohner et al., 2020; Cheah et al., 2021; Sheikh et al., 2015, 2017).

Bioactive glasses (BG) are a group of silicate-based synthetic alloplastic reactive materials with a unique capacity to form bonds with mineralized hard tissues such as bone in physiological environments (Roca-Millan et al., 2022). When BGs come into contact with body fluid, two interconnected phases of biological activity begin: a chemical exchange mechanism involving the formation of a hydroxycarbonate apatite (HCA) layer on the BG surface, and a cellular mechanism guiding osteogenesis (Jones, 2015).

During the chemical exchange process, a rapid ion exchange occurs between cations (H^+) in body fluids and sodium and potassium ions within the BG; silanol bonds ($Si-OH$) form during this process. The accumulating silanol groups raise the pH of the environment and trigger further chemical reactions in the silica glass network of the BG, increasing additional silanol release. The silanol groups then condense to form a silica-rich layer; calcium and phosphate ions migrating from the extracellular fluid accumulate in this layer (Jones, 2015; Välimäki & Aro, 2006). Following completion of the chemical exchange phase, an appropriate HCA

layer for bone formation takes shape; bone-forming progenitor cells and other associated cells adhere to this layer and differentiate to form bone matrix (Cheah et al., 2021).

Calcium compounds constitute one of the most promising groups among synthetic bone substitute materials, with calcium phosphate (CP) and calcium sulfate (CS) cements being prominent within this group. CP cement plays important roles in cell adhesion and tissue formation by affecting the adsorption of extracellular matrix proteins on its surface (Tsapikouni & Missirlis, 2008). Calcium stimulates osteoblastic bone synthesis, prolongs the lifespan of osteoblasts, and regulates the formation and resorptive function of osteoclasts (Kuroda et al., 2008). Phosphate regulates the differentiation and growth of osteoblasts and osteoblastic lineage cells; in addition, it suppresses osteoclast differentiation and bone resorption by modulating the RANK-ligand:OPG ratio (Zhang et al., 2011).

Despite their various clinical advantages, alloplastic materials have important limitations. The most fundamental disadvantage of these materials is that they lack osteogenesis and osteoinduction capacity; therefore, they cannot initiate new bone formation on their own and serve only as an osteoconductive scaffold (Ferraz, 2023). Furthermore, alloplastic materials are insufficient on their own in large-volume bone defects; in such cases, combination with autograft, allograft, or growth factors is required (Ferraz, 2023).

Factors Affecting Osteogenesis

To achieve a clinically successful bone graft, the material must meet certain properties. An ideal bone substitute material should be biocompatible, bioresorbable, osteoconductive, osteoinductive, structurally similar to bone, easy to use, and cost-effective (Kolk et al., 2012). Porosity is one of the primary

determinants of successful bone integration. Interconnected pores with a diameter exceeding 100 μm allow for cell infiltration, bone growth, and vascularization (Cornell, 1999; Klawitter & Weinstein, 1974), while pore sizes in the range of 150–500 μm are considered optimal for interface engineering activities, bone penetration, and implant absorption (Hertz & Bruce, 2007). High porosity and low density provide a larger surface area for vascularization and bone ingrowth. Vascularization is also a critical factor that directly affects the osteogenesis process. In cases where resorption and new bone formation do not occur simultaneously, biomechanically insufficient connective tissue formation may occur (Kao & Scott, 2007). Growth factors, particularly bone morphogenetic proteins (BMPs), are among the most powerful regulators of the osteogenesis process. BMPs initiate new bone formation by stimulating the differentiation of mesenchymal stem cells into osteoblasts. The inadequacy of carrier systems in osteoinductive agents and their short half-lives constitute the main limitations in clinical application (Kolk et al., 2012).

Clinical Applications

Bone grafts are routinely applied in many areas of dentistry, primarily dental implant placement, alveolar ridge augmentation, sinus floor elevation, socket preservation, and periodontal surgery (Ferraz, 2023; Zhao et al., 2021).

In dental implant applications, sufficient bone volume and density are of decisive importance for the long-term success of the implant. When there is insufficient bone, bone grafting becomes necessary prior to or simultaneously with implant placement (Zhao et al., 2021).

Ridge augmentation procedures are performed to reconstruct the height and width of the atrophic alveolar bone crest. This procedure can be planned to provide a suitable foundation for dental

implant placement or to improve gingival appearance (R et al., 2022).

When there is insufficient bone height in the posterior maxilla, sinus floor elevation procedures are applied, in which the sinus membrane is lifted and bone graft material is placed between the sinus membrane and the alveolar bone (Zhao et al., 2021). Various graft substitutes including autograft, allograft, xenograft, and alloplastic materials can be used in this procedure (Albadani et al., 2024).

Socket preservation procedures are applied to minimize dimensional changes in the alveolar bone following tooth extraction and to preserve bone volume for future dental implant placement. By placing graft material into the extraction socket, the bone structure is preserved and resorption is significantly reduced (Ferraz, 2023).

In advanced periodontal bone loss, bone grafts constitute an integral part of periodontal regeneration procedures. In these applications, the goals include filling periodontal defects and reconstructing alveolar bone (Ferraz, 2023; Zhao et al., 2021).

Conclusion

Bone graft materials play an indispensable role in the repair of bone defects in dentistry and oral-maxillofacial surgery. Each of the autograft, allograft, xenograft, and alloplastic materials contributes to bone regeneration at different levels through the mechanisms of osteogenesis, osteoinduction, and osteoconduction. Despite all their superior biological properties, the search for alternatives to autograft continues due to disadvantages such as donor site morbidity and limited material quantity. Alloplastic materials are expected to offer more predictable clinical outcomes with increasingly advancing bioengineering technologies. The decisive effects of porosity, vascularization, and growth factors on

osteogenesis directly guide clinical decision-making processes in material selection. Bone grafts, which are effectively used in a wide range of clinical applications including dental implants, sinus floor elevation, ridge augmentation, socket preservation, and periodontal regeneration, continue to be the primary determinant in achieving successful clinical outcomes with evidence-based selection criteria and individualized treatment protocols.

REFERENCES

- Albadani, M. M., Elayah, S. A., Al-Wesabi, M. A., Al-Aroomi, O. A., Al Qadasy, N. E. & Saleh, H. (2024). A graftless maxillary sinus lifting approach with simultaneous dental implant placement: a prospective clinical study. *BMC Oral Health*, 24(1), 227. <https://doi.org/10.1186/s12903-024-03949-9>
- Altıparmak, N., Soydan, S. S. & Uçkan, S. (2015). The effect of conventional surgery and piezoelectric surgery bone harvesting techniques on the donor site morbidity of the mandibular ramus and symphysis. *International Journal of Oral and Maxillofacial Surgery*, 44(9), 1131–1137. <https://doi.org/10.1016/j.ijom.2015.04.009>
- Amid, R., Kheiri, A., Kheiri, L., Kadkhodazadeh, M. & Ekhlasmandkermani, M. (2021). Structural and chemical features of xenograft bone substitutes: A systematic review of in vitro studies. *Biotechnology and Applied Biochemistry*, 68(6), 1432–1452. <https://doi.org/10.1002/bab.2065>
- Baldwin, P., Li, D. J., Auston, D. A., Mir, H. S., Yoon, R. S. & Koval, K. J. (2019). Autograft, Allograft, and Bone Graft Substitutes: Clinical Evidence and Indications for Use in the Setting of Orthopaedic Trauma Surgery. *Journal of Orthopaedic Trauma*, 33(4), 203–213. <https://doi.org/10.1097/BOT.0000000000001420>
- Bohner, M., Santoni, B. L. G. & Döbelin, N. (2020). β -tricalcium phosphate for bone substitution: Synthesis and properties. *Acta Biomaterialia*, 113, 23–41. <https://doi.org/10.1016/j.actbio.2020.06.022>
- Chappard, D. (2017). Beta-tricalcium phosphate and bone surgery: Editorial. *Morphologie : Bulletin de l'Association Des Anatomistes*, 101(334), 111–112. <https://doi.org/10.1016/j.morpho.2017.09.001>
- Cheah, C. W., Al-Namnam, N. M., Lau, M. N., Lim, G. S., Raman, R., Fairbairn, P. & Ngeow, W. C. (2021). Synthetic Material for Bone, Periodontal, and Dental

Tissue Regeneration: Where Are We Now, and Where Are We Heading Next? *Materials (Basel, Switzerland)*, 14(20). <https://doi.org/10.3390/ma14206123>

Ciszyński, M., Dominiak, S., Dominiak, M., Gedrange, T. & Hadzik, J. (2023). Allogenic Bone Graft in Dentistry: A Review of Current Trends and Developments. *International Journal of Molecular Sciences*, 24(23). <https://doi.org/10.3390/ijms242316598>

Cornell, C. N. (1999). Osteoconductive materials and their role as substitutes for autogenous bone grafts. *The Orthopedic Clinics of North America*, 30(4), 591–598. [https://doi.org/10.1016/s0030-5898\(05\)70112-7](https://doi.org/10.1016/s0030-5898(05)70112-7)

Develioglu, H., Unver Saraydin, S. & Kartal, U. (2009). The bone-healing effect of a xenograft in a rat calvarial defect model. *Dental Materials Journal*, 28(4), 396–400. <https://doi.org/10.4012/dmj.28.396>

Ehrler, D. M. & Vaccaro, A. R. (2000). The use of allograft bone in lumbar spine surgery. *Clinical Orthopaedics and Related Research*, 371, 38–45. <https://doi.org/10.1097/00003086-200002000-00005>

El-Rashidy, A. A., Roether, J. A., Harhaus, L., Kneser, U. & Boccaccini, A. R. (2017). Regenerating bone with bioactive glass scaffolds: A review of in vivo studies in bone defect models. *Acta Biomaterialia*, 62, 1–28. <https://doi.org/10.1016/j.actbio.2017.08.030>

Elsalanty, M. E. & Genecov, D. G. (2009). Bone grafts in craniofacial surgery. *Cranio-maxillofacial Trauma & Reconstruction*, 2(3), 125–134. <https://doi.org/10.1055/s-0029-1215875>

Fan, Y.-P., Lu, J.-F., Xu, A.-T. & He, F.-M. (2018). Physiochemical characterization and biological effect of anorganic bovine bone matrix and organic-containing bovine bone matrix in comparison with Bio-Oss in rabbits. *Journal of Biomaterials Applications*, 33(4), 566–575. <https://doi.org/10.1177/0885328218804926>

Ferraz, M. P. (2023). Bone Grafts in Dental Medicine: An Overview of Autografts, Allografts and Synthetic Materials. *Materials (Basel, Switzerland)*, 16(11). <https://doi.org/10.3390/ma16114117>

Frayssinet, P., Fages, J., Bonel, G. & Rouquet, N. (1998). Biotechnology, material sciences and bone repair. *European Journal of Orthopaedic Surgery & Traumatology*, 8(1), 17–25. <https://doi.org/10.1007/bf01782892>

Goldberg, V. M. & Stevenson, S. (1987). Natural history of autografts and allografts. *Clinical Orthopaedics and Related Research*, 225, 7–16. <https://www.ncbi.nlm.nih.gov/pubmed/3315383>

Greenwald, A. S., Boden, S. D., Goldberg, V. M., Khan, Y., Laurencin, C. T., Rosier, R. N. & American Academy of Orthopaedic Surgeons. The Committee on Biological Implants. (2001). Bone-graft substitutes: facts, fictions, and applications. *The Journal of Bone and Joint Surgery. American Volume*, 83-A Suppl 2 Pt 2, 98–103. <https://doi.org/10.2106/00004623-200100022-00007>

Haugen, H. J., Sanz, J., Perale, G., Saiz, A. M., Romandini, M. & Rahmati, M. (2026). Bone Grafts: Everything You Need to Know. *Journal of Periodontal Research*. <https://doi.org/10.1111/jre.70087>

Heilmann, F., Standard, O. C., Müller, F. A. & Hoffman, M. (2007). Development of graded hydroxyapatite/CaCO(3) composite structures for bone ingrowth. *Journal of Materials Science. Materials in Medicine*, 18(9), 1817–1824. <https://doi.org/10.1007/s10856-007-3028-3>

Hench, L. L. (1991). Bioceramics: From concept to clinic. *Journal of the American Ceramic Society. American Ceramic Society*, 74(7), 1487–1510. <https://doi.org/10.1111/j.1151-2916.1991.tb07132.x>

Hertz, A. & Bruce, I. J. (2007). Inorganic materials for bone repair or replacement applications. *Nanomedicine (London, England)*, 2(6), 899–918. <https://doi.org/10.2217/17435889.2.6.899>

Jones, J. R. (2015). Reprint of: Review of bioactive glass: From Hench to hybrids. *Acta Biomaterialia*, 23 Suppl, S53–S82. <https://doi.org/10.1016/j.actbio.2015.07.019>

Jung, Y., Kim, W.-H., Lee, S.-H., Ju, K. W., Jang, E.-H., Kim, S.-O., Kim, B. & Lee, J.-H. (2020). Evaluation of New Octacalcium Phosphate-Coated Xenograft in Rats Calvarial Defect Model on Bone Regeneration. *Materials (Basel, Switzerland)*, 13(19). <https://doi.org/10.3390/ma13194391>

Kämmerer, P. W. & Al-Nawas, B. (2023). Bone reconstruction of extensive maxillomandibular defects in adults. *Periodontology 2000*, 93(1), 340–357. <https://doi.org/10.1111/prd.12499>

Kao, S. T. & Scott, D. D. (2007). A review of bone substitutes. *Oral and Maxillofacial Surgery Clinics of North America*, 19(4), 513–521, vi. <https://doi.org/10.1016/j.coms.2007.06.002>

Keating, J. F. & McQueen, M. M. (2001). Substitutes for autologous bone graft in orthopaedic trauma. *The Journal of Bone and Joint Surgery. British Volume*, 83(1), 3–8. <https://doi.org/10.1302/0301-620x.83b1.11952>

Khan, S. N., Cammisia, F. P., Jr, Sandhu, H. S., Diwan, A. D., Girardi, F. P. & Lane, J. M. (2005). The biology of bone grafting. *The Journal of the American Academy of Orthopaedic Surgeons*, 13(1), 77–86. <https://www.ncbi.nlm.nih.gov/pubmed/15712985>

Klawitter, J. J. & Weinstein, A. M. (1974). The status of porous materials to obtain direct skeletal attachment by tissue ingrowth. *Acta Orthopaedica Belgica*, 40(5-6), 755–765. <https://www.ncbi.nlm.nih.gov/pubmed/4469737>

Kolk, A., Handschel, J., Drescher, W., Rothamel, D., Kloss, F., Blessmann, M., Heiland, M., Wolff, K.-D. & Smeets, R. (2012). Current trends and future perspectives of bone substitute materials - from space holders to innovative biomaterials. *Journal of Cranio-Maxillo-Facial Surgery : Official Publication of the European Association for Cranio-Maxillo-Facial Surgery*, 40(8), 706–718. <https://doi.org/10.1016/j.jcms.2012.01.002>

Kübler, A., Neugebauer, J., Oh, J.-H., Scheer, M. & Zöller, J. E. (2004). Growth and proliferation of human osteoblasts on different bone graft substitutes: an in vitro study. *Implant Dentistry*, 13(2), 171–179. <https://doi.org/10.1097/01.id.0000127522.14067.11>

Kuroda, Y., Hisatsune, C., Nakamura, T., Matsuo, K. & Mikoshiba, K. (2008). Osteoblasts induce Ca²⁺ oscillation-independent NFATc1 activation during osteoclastogenesis. *Proceedings of the National Academy of Sciences of the United States of America*, 105(25), 8643–8648. <https://doi.org/10.1073/pnas.0800642105>

Long, B., Dan, L., Jian, L., Yunyu, H., Shu, H. & Zhi, Y. (2012). Evaluation of a novel reconstituted bone xenograft using processed bovine cancellous bone in combination with purified bovine bone morphogenetic protein. *Xenotransplantation*, 19(2), 122–132. <https://doi.org/10.1111/j.1399-3089.2012.00694.x>

Lu, H., Zhou, Y., Ma, Y., Xiao, L., Ji, W., Zhang, Y. & Wang, X. (2021). Current application of beta-tricalcium phosphate in bone repair and its mechanism to regulate osteogenesis. *Frontiers in Materials*, 8. <https://doi.org/10.3389/fmats.2021.698915>

Martinetti, R., Dolcini, L. & Mangano, C. (2005). Physical and chemical aspects of a new porous hydroxyapatite. *Analytical and Bioanalytical Chemistry*, 381(3), 634–638. <https://doi.org/10.1007/s00216-004-2957-7>

Marx, R. E. (2007). Bone and bone graft healing. *Oral and Maxillofacial Surgery Clinics of North America*, 19(4), 455–466, v. <https://doi.org/10.1016/j.coms.2007.07.008>

Miron, R. J. (2024). Optimized bone grafting. *Periodontology 2000*, 94(1), 143–160. <https://doi.org/10.1111/prd.12517>

Nandi, S. K., Roy, S., Mukherjee, P., Kundu, B., De, D. K. & Basu, D. (2010). Orthopaedic applications of bone graft & graft substitutes: a review. *The Indian Journal of Medical Research*, 132, 15–30. <https://www.ncbi.nlm.nih.gov/pubmed/20693585>

Oryan, A., Alidadi, S., Moshiri, A. & Maffulli, N. (2014). Bone regenerative medicine: classic options, novel strategies, and future directions. *Journal of Orthopaedic Surgery and Research*, 9(1), 18. <https://doi.org/10.1186/1749-799X-9-18>

Oryan, A., Meimandi Parizi, A., Shafiei-Sarvestani, Z. & Bigham, A. S. (2012). Effects of combined hydroxyapatite and human platelet rich plasma on bone healing in rabbit model: radiological, macroscopical, histopathological and biomechanical evaluation. *Cell and Tissue Banking*, 13(4), 639–651. <https://doi.org/10.1007/s10561-011-9285-x>

Oryan, A., Moshiri, A., Meimandi Parizi, A. H. & Raayat Jahromi, A. (2012). Repeated administration of exogenous Sodium-hyaluronate improved tendon healing in an in vivo transection model. *Journal of Tissue Viability*, 21(3), 88–102. <https://doi.org/10.1016/j.jtv.2012.06.002>

Pereira-Júnior, O. C. M., Rahal, S. C., Iamaguti, P., Felisbino, S. L., Pavan, P. T. & Vulcano, L. C. (2007). Comparison between polyurethanes containing castor oil (soft segment) and cancellous bone autograft in the treatment of segmental bone defect induced in rabbits. *Journal of Biomaterials Applications*, 21(3), 283–297. <https://doi.org/10.1177/0885328206063526>

R, A., Koduganti, R. R., Harika, T. S. L. & Rajaram, H. (2022). Ridge Augmentation Is a Prerequisite for Successful Implant Placement: A Literature Review. *Cureus*, 14(1), e20872. <https://doi.org/10.7759/cureus.20872>

Roberts, T. T. & Rosenbaum, A. J. (2012). Bone grafts, bone substitutes and orthobiologics: the bridge between basic science and clinical advancements in fracture healing. *Organogenesis*, 8(4), 114–124. <https://doi.org/10.4161/org.23306>

Roca-Millan, E., Jané-Salas, E., Marí-Roig, A., Jiménez-Guerra, Á., Ortiz-García, I., Velasco-Ortega, E., López-López, J. & Monsalve-Guil, L. (2022). The Application of Beta-Tricalcium Phosphate in Implant Dentistry: A Systematic Evaluation of Clinical Studies. *Materials (Basel, Switzerland)*, 15(2). <https://doi.org/10.3390/ma15020655>

Serrano Méndez, C. A., Lang, N. P., Caneva, M., Ramírez Lemus, G., Mora Solano, G. & Botticelli, D. (2017). Comparison of allografts and xenografts used for alveolar ridge preservation. A clinical and histomorphometric RCT in humans. *Clinical Implant Dentistry and Related Research*, 19(4), 608–615. <https://doi.org/10.1111/cid.12490>

Sheikh, Z., Hamdan, N., Ikeda, Y., Gryn timer, M., Ganss, B. & Glogauer, M. (2017). Natural graft tissues and synthetic biomaterials for periodontal and alveolar bone reconstructive applications: a review. *Biomaterials Research*, 21, 9. <https://doi.org/10.1186/s40824-017-0095-5>

Sheikh, Z., Najeeb, S., Khurshid, Z., Verma, V., Rashid, H. & Glogauer, M. (2015). Biodegradable Materials for Bone Repair and Tissue Engineering Applications. *Materials (Basel, Switzerland)*, 8(9), 5744–5794. <https://doi.org/10.3390/ma8095273>

Stevens, M. R., Ghasemi, S. & Tabrizi, R. (2021). *Innovative Perspectives in Oral and Maxillofacial Surgery*. Springer Nature. <https://play.google.com/store/books/details?id=swM7EAAAQBAJ>

Supová, M. (2009). Problem of hydroxyapatite dispersion in polymer matrices: a review. *Journal of Materials Science. Materials in Medicine*, 20(6), 1201–1213. <https://doi.org/10.1007/s10856-009-3696-2>

Thieu, M. K. L., Haugen, H. J., Sanz-Esporrin, J., Sanz, M., Lyngstadaas, S. P. & Verket, A. (2021). Guided bone regeneration of chronic non-contained bone defects using a volume stable porous block TiO₂ scaffold: An experimental in vivo study. *Clinical Oral Implants Research*, 32(3), 369–381. <https://doi.org/10.1111/clr.13708>

Tsapikouni, T. S. & Missirlis, Y. F. (2008). Protein–material interactions: From micro-to-nano scale. *Materials Science & Engineering. B, Solid-State Materials for Advanced Technology*, 152(1-3), 2–7. <https://doi.org/10.1016/j.mseb.2008.06.007>

Turhani, D., Item, C., Thurnher, D., Kapral, D., Cvikl, B., Weissenböck, M., Yerit, K., Erovic, B., Moser, D., Watzinger, F., Ewers, R. & Lauer, G. (2003). [Evidence of osteocalcin expression in osteoblast cells of mandibular origin growing on biomaterials with RT-PCR and SDS-PAGE/Western blotting]. *Mund-, Kiefer- und Gesichtschirurgie : MKG*, 7(5), 294–300. <https://doi.org/10.1007/s10006-003-0495-7>

Turhani, D., Weissenböck, M., Watzinger, E., Yerit, K., Cvikl, B., Ewers, R. & Thurnher, D. (2005). Invitro study of adherent mandibular osteoblast-like cells on carrier materials. *International Journal of Oral and Maxillofacial Surgery*, 34(5), 543–550. <https://doi.org/10.1016/j.ijom.2004.10.023>

Välimäki, V. V. & Aro, H. T. (2006). Molecular basis for action of bioactive glasses as bone graft substitute. *Scandinavian Journal of Surgery : SJS : Official Organ for the Finnish Surgical Society and the Scandinavian Surgical Society*, 95(2), 95–102. <https://doi.org/10.1177/145749690609500204>

Zhang, R., Lu, Y., Ye, L., Yuan, B., Yu, S., Qin, C., Xie, Y., Gao, T., Drezner, M. K., Bonewald, L. F. & Feng, J. Q. (2011). Unique roles of phosphorus in endochondral bone formation and osteocyte maturation. *Journal of Bone and Mineral Research : The Official Journal of the American Society for Bone and Mineral Research*, 26(5), 1047–1056. <https://doi.org/10.1002/jbmr.294>

Zhao, R., Yang, R., Cooper, P. R., Khurshid, Z., Shavandi, A. & Ratnayake, J. (2021). Bone Grafts and Substitutes in Dentistry: A Review of Current Trends and Developments. *Molecules (Basel, Switzerland)*, 26(10). <https://doi.org/10.3390/molecules26103007>

Zimmermann, G. & Moghaddam, A. (2011). Allograft bone matrix versus synthetic bone graft substitutes. *Injury*, 42 Suppl 2, S16–S21. <https://doi.org/10.1016/j.injury.2011.06.199>

